

This Provisional PDF corresponds to the article as it appeared upon acceptance. Fully formatted PDF and full text (HTML) versions will be made available soon.

## Niemann-Pick disease type C

*Orphanet Journal of Rare Diseases* 2010, **5**:16 doi:10.1186/1750-1172-5-16

Marie T Vanier (marie-t.vanier@inserm.fr)

**ISSN** 1750-1172

**Article type** Review

**Submission date** 7 October 2009

**Acceptance date** 3 June 2010

**Publication date** 3 June 2010

**Article URL** <http://www.ojrd.com/content/5/1/16>

This peer-reviewed article was published immediately upon acceptance. It can be downloaded, printed and distributed freely for any purposes (see copyright notice below).

Articles in *Orphanet Journal of Rare Diseases* are listed in PubMed and archived at PubMed Central.

For information about publishing your research in *Orphanet Journal of Rare Diseases* or any BioMed Central journal, go to

<http://www.ojrd.com/info/instructions/>

For information about other BioMed Central publications go to

<http://www.biomedcentral.com/>

## **Niemann-Pick disease type C**

**Marie T Vanier<sup>1,2</sup>**

<sup>1</sup>Institut National de la Santé et de la Recherche Médicale, Unité 820, Faculté de Médecine Lyon-Est Claude Bernard, 7 Rue G. Paradin, F-69008, Lyon, France ; <sup>2</sup>Hospices Civils de Lyon, Laboratoire de Neurobiologie Gillet-Mérieux, Centre de Biologie Est, 59 Bd Pinel, F-69500, Bron, France

E-mails : [marie-t.vanier@inserm.fr](mailto:marie-t.vanier@inserm.fr) or [vanier@attglobal.net](mailto:vanier@attglobal.net)

## Abstract

Niemann-Pick C disease (NP-C) is a neurovisceral atypical lysosomal lipid storage disorder with an estimated minimal incidence of 1/120 000 live births. The broad clinical spectrum ranges from a neonatal rapidly fatal disorder to an adult-onset chronic neurodegenerative disease. The neurological involvement defines the disease severity in most patients but is typically preceded by systemic signs (cholestatic jaundice in the neonatal period or isolated spleno- or hepatosplenomegaly in infancy or childhood). The first neurological symptoms vary with age of onset: delay in developmental motor milestones (early infantile period), gait problems, falls, clumsiness, cataplexy, school problems (late infantile and juvenile period), and ataxia not unfrequently following initial psychiatric disturbances (adult form). The most characteristic sign is vertical supranuclear gaze palsy. The neurological disorder consists mainly of cerebellar ataxia, dysarthria, dysphagia, and progressive dementia. Cataplexy, seizures and dystonia are other common features. NP-C is transmitted in an autosomal recessive manner and is caused by mutations of either the *NPC1* (95% of families) or the *NPC2* genes. The exact functions of the NPC1 and NPC2 proteins are still unclear. NP-C is currently described as a cellular cholesterol trafficking defect but in the brain, the prominently stored lipids are gangliosides. Clinical examination should include comprehensive neurological and ophthalmological evaluations. The primary laboratory diagnosis requires living skin fibroblasts to demonstrate accumulation of unesterified cholesterol in perinuclear vesicles (lysosomes) after staining with filipin. Pronounced abnormalities are observed in about 80% of the cases, mild to moderate alterations in the remainder ("variant" biochemical phenotype). Genotyping of patients is useful to confirm the diagnosis in the latter patients and essential for future prenatal diagnosis. The differential diagnosis may include other lipidoses; idiopathic neonatal hepatitis and other causes of cholestatic icterus should be considered in neonates, and conditions with cerebellar ataxia, dystonia, cataplexy and supranuclear gaze palsy in older children and adults. Symptomatic management of patients is crucial. A first product, miglustat, has been granted marketing authorization in Europe and several other countries for specific treatment of the neurological manifestations. The prognosis largely correlates with the age at onset of the neurological manifestations.

## Disease definition

### *Historical delineation*

Coined in the late 1920's from the pioneering work of Albert Niemann and Ludwig Pick, the eponym "Niemann-Pick disease" has since been used to designate a heterogeneous group of autosomal recessive lysosomal lipid storage disorders, with common features of hepatosplenomegaly and sphingomyelin storage in reticuloendothelial and parenchymal tissues, with or without neurological involvement. In 1958, Crocker and Farber showed that there was a wide variability in age of onset and clinical expression, as well as in the level of sphingomyelin storage in tissues [1]. This led Crocker to propose a classification into four subgroups, A to D [2]. Type A was characterized by severe, early CNS deterioration and massive visceral and cerebral sphingomyelin storage. Type B showed a chronic course with marked visceral involvement but a sparing of the nervous system. Types C and D were characterized by a sub acute nervous system involvement with a moderate and slower course and a milder visceral storage. Type D patients were individualized essentially on their homogenous Nova Scotia Acadian origin. In 1966, Brady and associates [3] demonstrated a severe deficiency in sphingomyelinase activity in tissues from patients with type A, a finding soon extended to type B, but not to types C and D, indicating that the two latter types constituted separate entities. From that time on, with a turn following seminal observations in a Balb/c murine model of the disorder [4], the concept of Niemann-Pick type C disease evolved from that of a sphingomyelin storage disorder to that of a cholesterol storage disorder [5]. This and later work led to the reclassification of type C as a cellular lipid trafficking disorder, involving more specially, but not only, endocytosed cholesterol.

### *Definition of Niemann-Pick disease type C*

Today, by definition, "Niemann-Pick C disease" encompasses disorders characterized by unique abnormalities of intracellular transport of endocytosed cholesterol with sequestration of unesterified cholesterol in lysosomes and late endosomes [5-12]. Major advances have been the description of two genetic complementation groups [13,14] and the subsequent isolation of the two underlying genes [15,16]. *NPC1* is involved in 95% of the families [14], including those with type D [17]. *NPC2* is involved in rare families (about 30 are known to date). Although the precise functions of the NPC1 and NPC2 proteins are still elusive, current knowledge supports the idea that these proteins function in a coordinate fashion and that they are involved in the cellular postlysosomal/late endosomal transport of cholesterol and other molecules [10-12, 18, 19].

Niemann-Pick diseases thus oppose two clearly distinct groups: acid sphingomyelinase deficiencies (due to *SMPD1* mutations, including types A, B and intermediate forms,) and Niemann-Pick type C, with alterations in trafficking of endocytosed cholesterol (due to *NPC1* or *NPC2* mutations). Type D as a distinct entity is no longer justified. From a practical standpoint, no patient should today be longer qualified of suffering from "Niemann-Pick disease" without specification of the subgroup, either acid sphingomyelinase deficiency or type C.

## Disease name and synonyms

"Niemann-Pick disease type C" (or "Niemann-Pick C disease"), often abbreviated as NP-C (or NPC), is currently the generic name widely used to designate the condition, irrespective of which gene, *NPC1* or *NPC2*, is mutated. This term now encompasses the historical Niemann-Pick disease type D referring to the "Nova Scotia" isolate, later shown to be a genetic *NPC1*

variant [17]. Instead, a subdivision is sometimes made between Niemann-Pick C1 (NP-C1) or C2 (NP-C2), according to the gene involved. Patients with a retrospective diagnosis of Niemann-Pick C disease have also been described in the 1960s and 1970s as juvenile Niemann-Pick disease, juvenile dystonic lipidosis, atypical cerebral lipidosis, neurovisceral storage disease with vertical supranuclear ophthalmoplegia, maladie de Neville, DAF (down-gaze paresis, ataxia, foam cell) syndrome, adult dystonic lipidosis, adult neurovisceral lipidosis, giant cell hepatitis, and lactosylceramidosis [10,20].

## Epidemiology

NP-C (either NP-C1 or NP-C2) shows autosomal recessive inheritance and is panethnic. The true prevalence of NP-C is difficult to assess, because of insufficient clinical awareness combined with the relative difficulty of biochemical testing. Estimates of birth prevalence ranging between 0.66 and 0.83 per 100,000 were proposed for France, the UK and Germany based on diagnoses made in the laboratory of the author over the period 1988-2002 [10, 11]. However, very different figures of 0.47, 0.35 and 2.20 per 100,000, respectively, were reported in studies from Australia (20 cases between 1980-1996), The Netherlands (25 cases between 1970-1996) and Northern Portugal (9 cases 1985-2003) [21-23]. The low incidence found for Australia and the Netherlands might be explained by a non-exhaustivity of the diagnoses in the years of birth of many patients. The wide clinical spectrum of NP-C was not recognized until the early 1990s, especially regarding rapidly fatal infantile cases, and no specific laboratory testing was available until the mid 1980s. For this review, an updated incidence of 0.82/100,000 was calculated for France, considering the total number of cases (n=63) diagnosed for French hospitals during the 2000-2009 period vs. the number of births during the same period, a possibly more appropriate way of calculation. This value should be considered as a minimal estimate, since atypical phenotypes may not be suspected clinically or may be missed by the diagnostic laboratory. Including prenatal cases from terminated pregnancies during the same period (n=11) increased the incidence to 0.96 per 100,000.

Most families (about 95%) belong to the NP-C1 group. Two NP-C1 isolates have been described. The first one, in French Acadians originating from Normandy and originally established in Nova Scotia, was initially described as Niemann-Pick disease type D; it is characterized by the *NPC1* p.G992W mutation [1, 17, and 24]. Another isolate was described in Hispanics from southern Colorado and New Mexico with their roots in the Upper Rio Grande valley of the USA, carrying the *NPC1* p. I1061T mutation [25, 26].

## Clinical Description

The clinical presentation of NP-C is extremely heterogeneous, with an age of onset ranging from the perinatal period until well into adult age (as late as the seventh decade of life). Similarly, the lifespan of the patients varies between a few days until over 60 years of age, although a majority of cases die between 10 and 25 years of age [10,11,27-30]. The clinical spectrum discussed below has been analyzed from several large surveys [28-35].

NP-C is classically a neurovisceral condition. Importantly, visceral involvement (of liver, spleen, and sometimes lung) and neurologic or psychiatric manifestations arise at different times, and they also follow completely independent courses. Apart from a small subset of patients who die at birth, or in the first 6 months of life from hepatic or respiratory failure, and exceptional adult cases, all patients ultimately will develop a progressive and fatal neurological disease. Systemic disease, when present, always precedes onset of neurological

symptoms, but the systemic component may be absent or minimal in approximately 15% of all patients, and close to half of the adult-onset patients, at least at the time of diagnosis. In typical patients, the neurologic disorder consists mainly of cerebellar ataxia, dysarthria, dysphagia, and progressive dementia, and the majority of cases show a characteristic vertical supranuclear gaze palsy (VSGP) [36]. Cataplexy, seizures, and dystonia are other quite common features, and psychiatric disturbances are frequent in late-onset patients. The proper recognition of VSGP is essential but this sign is often overlooked at an early stage, because slow pursuit is often maintained although saccade velocity is already impaired. Cataplexy (with or without narcolepsy), usually laughter-induced, is another more specific symptom [37,38]. Except for the perinatal period, the systemic disease is usually not very severe and is well tolerated. The splenomegaly has been described to fluctuate and to decrease with time. Severe lung involvement has been reported in a few patients but is not frequent.

A description of the various clinical forms by age categories has been used in recent reviews [10,11,27] and will also be followed in this summary. Detailed complementary information can be obtained in [27]. For each age category except for the perinatal presentations, one should however distinguish patients entering the disease by systemic involvement [39] from those who are starting then their neurological disease (although they may have presented earlier with visceral symptoms). Of essential importance is to note that the age of onset of the systemic symptoms is not related with that of the neurological disease (the latter can occur many years or even decades later), while there is a correlation between the age of onset of the neurological symptoms and the general further course of the disease and lifespan (Fig. 1). Categorizing patients by forms based on the age range of onset of the neurological symptoms [11,32,40], irrespective of the age of the first symptom, is very useful for genetic counseling, natural history studies and also in clinical practice. With an exception for the severe early infantile neurological form which is quite significantly distinct, recent large studies have however demonstrated an overlap between the neurological forms, and thus a continuum [27]. A schematic representation is proposed in Fig. 2.

### ***Perinatal presentation***

Niemann-Pick C disease is now recognized as a relatively common cause of liver disease in early life. Fetal hydrops or fetal ascites can be observed [28]. Above all, a prolonged neonatal cholestatic icterus, appearing in the first days or weeks of life and usually associated with progressive hepatosplenomegaly is present in close to half of patients, although with very variable intensity [29,41,42]. In most cases, the icterus resolves spontaneously by 2 to 4 months of age, and only hepatosplenomegaly remains for a highly variable period, preceding onset of neurologic symptoms (see below). In about 10% of these patients, however, the icterus quickly worsens and leads to liver failure. Children with this dramatic "acute" neonatal cholestatic rapidly fatal form usually die before the age of 6 months [29]. Some other infants, especially (but not exclusively) those having mutations in the *NPC2* gene, present with a severe respiratory insufficiency (together with hepatosplenomegaly or more severe liver disease) that may also be fatal. In two patients, lung lavage, radiology and histology showed signs of pulmonary alveolar lipoproteinosis [43,44]. Patients with NP-C do not show neurological manifestations during the neonatal period (important for differential diagnosis). But there are many examples of patients dying from a severe perinatal form having siblings with a neurologic infantile or juvenile onset form [11,29].

### ***Early infantile period (2 months-2 years)***

#### ***Systemic stage***

An isolated hepatosplenomegaly can be discovered at this age period, which may well stay

isolated for many years, in spite of the early onset. Once the diagnosis of NP-C is made, a regular neuropsychiatric follow up should be initiated.

*Severe early infantile neurologic onset form.*

In these infants, hepatosplenomegaly has almost invariably been present since birth or the first months of life. Delay of developmental motor milestones from the age of 8-9 months and central hypotonia constitute the first neurologic symptoms, which become evident between the age of 1 and 2 years. Subsequent clinical course includes a loss of acquired motor skills, proportionally less marked mental regression, followed by pronounced spasticity with pyramidal tract involvement. Many of these children never learn to walk. Intention tremor is frequently present; supranuclear gaze palsy is usually not recognized. Seizures are uncommon. Brain imaging (MRI and MRS) shows signs of leukodystrophy and cerebral atrophy. Survival rarely exceeds 5 years. This form seems to be more frequent in Southern Europe (where it constitutes >20% of the cases) and the Middle East [11,29,32].

***Late infantile period (2 to 6 years)***

*Systemic stage*

Many patients start their disease by discovery of an isolated hepatosplenomegaly or splenomegaly during this period. Regular neuropsychiatric follow-up should be initiated, as above.

*Late infantile neurologic onset form*

Hepatosplenomegaly has almost invariably been present for a varying length of time. Language delay is frequent. The child often presents with gait problems, frequent falls and clumsiness between 3 and 5 years of age, due to ataxia. VSGP is usually present but may not be recognized at an early stage. Hearing loss has also been described. Cataplexy develops relatively frequently and may occasionally be the presenting symptom. The motor problems worsen, and impairment in mental development becomes more obvious. A significant proportion of patients develop seizures which may be partial, generalized, or both. They generally respond to standard treatment but refractory cases may occur, with some patients dying from status epilepticus or complications of seizures. Severe epilepsy has a bad prognosis and significantly shortens the lifespan of the patients. As ataxia progresses, dysphagia, dysarthria, and dementia develop. At later stages, the patients develop pyramidal signs and spasticity, and pronounced swallowing problems. Most require gastrostomy. Death most often occurs between 7 to 12 years in this form.

***Juvenile period (6-15 years) (classical form)***

*Systemic stage*

Discovery of an isolated splenomegaly (or, rarely, of a hepatosplenomegaly) at this period may again be the inaugural sign of the disease, and these patients should later be appropriately monitored.

*Juvenile neurologic onset form*

This constitutes in most countries the most common form of the disease. A moderate splenomegaly (or hepatosplenomegaly) is frequent, and may have been detected at any earlier time, including the neonatal period. However, cases in whom a splenomegaly had been noted in early childhood but is hardly detectable at the time first neurological symptoms arise are not rare; and absence of a detectable organomegaly has been reported to occur in at least 10% of cases. School problems with difficulties in writing and impaired attention are very common and may lead to misdiagnosis. The disease may also mimic dyspraxia. VSGP is almost

invariably present and often the initial sign. The child becomes clumsier and shows more learning disabilities. Cataplexy, with or without narcolepsy, typically laughter-induced, is another common symptom. Ataxia soon becomes obvious, with frequent falls and difficulties to run, and progresses at a variable rate. Dysarthria develops, as well as dysphagia. Action dystonia is also frequent, Motor impairment is major and intellectual decline may be variable. About half of the patients with the classic form develop seizures of variable type and severity (see above). At a later stage, dysarthria worsens and patients often stop talking. At a late stage, patients develop pyramidal signs and spasticity, and pronounced swallowing problems, requiring gastrostomy. The lifespan is quite variable, some patients being still alive by age 30 or later [27].

### ***Adolescent and adults (>15 years)***

#### *Systemic adult form of NP-C*

The finding of three patients aged 53-63 years with isolated splenomegaly and a biochemical and molecular diagnosis of NP-C [45-48] suggests the existence of a rare non-neuronopathic form of the disease (possibly corresponding to the ill-described historical "type E"). Nevertheless, apart from these exceptional cases and from infants with early death, as stated above, all NP-C patients develop neurologic symptoms.

#### *Adult neurologic onset form*

More patients with a neurologic adult onset form of the disease (often in the second or third decade, but as late as 50 years or older) have been described in recent years [30,35,49-53] This diagnosis is probably underestimated. Absence of clinically detectable splenomegaly has been reported in a significant proportion of patients but abdominal sonography may reveal a slightly enlarged spleen. VSGP is usually present but may also be missing. The most common symptomatology is that of an attenuated juvenile form with an insidious onset, with in at least one third of cases, a psychiatric presentation that may be isolated for several years before the onset of motor and cognitive signs. Psychiatric signs are most often consistent with psychosis, including paranoid delusions, auditory or visual hallucinations, and interpretative thoughts. Onset may be acute or progressive, eventually with relapses. At this stage the neurologic examination may be normal. Other types of psychiatric disturbances are depressive syndrome, behavioral problems with aggressiveness, or social isolation. Cases have also been reported with bipolar disorders, obsessive-compulsive disorders, or transient visual hallucinations. From compilation of the literature [35] the most common features are: cerebellar ataxia (76%), vertical supranuclear ophthalmoplegia (75%), dysarthria (63%), cognitive troubles (61%), movement disorders (58%), splenomegaly (54%), psychiatric disorders (45%) and dysphagia (37%). Movement disorders (dystonia, Parkinsonism, chorea) are more frequent than in the juvenile form. Some patients show severe ataxia, dystonia, and dysarthria with variable cognitive dysfunction, whereas psychiatric symptoms and dementia dominate in others. Epilepsy is rare in adult onset NP-C (15%). Later course is similar to that in the juvenile form.

## **Etiology**

Mutations in either of the two genes, *NPC1* or *NPC2*, may cause the disease [13-16]. Approximately 95% of patients have mutations in the *NPC1* gene, which encodes a large membrane glycoprotein with mostly a late endosomal localization [54]. The remainder have mutations in the *NPC2* gene, which encodes a small soluble lysosomal protein that binds cholesterol with high affinity [16,55,56]. Mutations in the *NPC1* or *NPC2* genes result in a

similar cellular lesion, including a unique impairment in processing and utilization of endocytosed cholesterol that could explain cholesterol storage and secondary alterations of sphingomyelin metabolism in extra neural tissues. Glycolipids and free sphingosine/sphinganine storage also occurs. In brain, - more specifically in neurons - the dominant lipid accumulation is in fact that of GM2 and GM3 gangliosides, with only limited apparent abnormalities of cholesterol (see below). Early studies in cells and tissues from NP-C1 and NP-C2 patients could not disclose any biochemical marker that was specific to any of the groups, suggesting that both proteins may function in tandem or sequentially [14]. Comparison between double mutant mice deficient in both NPC1 and NPC2 and the single mutants demonstrated a non-redundant functional cooperativity of the two proteins in a common pathway for lipid cellular transport, which strengthened this concept [18]. The exact functions of the NPC1 and NPC2 proteins are still unclear [10-12,56,57], which greatly complicates understanding of the pathophysiology. Neuronal storage with meganeurite formation and extensive growth of ectopic dendrites, as well as formation of neurofibrillary tangles, are important neuropathological features together with neuroinflammation and neuroaxonal dystrophy. As the disease progresses, neuronal death becomes prominent, affecting more specifically certain regions, particularly Purkinje cells of the cerebellum, but the basis of this selective neuronal vulnerability is still unknown [10,58].

#### ***Lipid accumulation in tissues.***

Similar profiles have been observed in NP-C1 and NP-C2 patients (and animal models), but the pattern of accumulating lipids is different in brain and in non-neural organs [10,18,20,40,59-64]. In liver and spleen, a complex pattern, with no predominating compound, is observed. Accumulated lipids include unesterified cholesterol and sphingomyelin (2- to 5-fold increase in human patients), bis(monoacylglycerol) phosphate (also named LBPA or BMP), glycolipids (essentially glucosylceramide and lactosylceramide), and free sphingosine and sphinganine. In human patients, the level of storage is more pronounced in the spleen than in the liver, where changes may be subtle. In brain tissue, neither cholesterol nor sphingomyelin overtly accumulate, but significant alterations of glycosphingolipids occur, especially for gangliosides GM2 and GM3 (10-20 fold increase). Free sphingosine levels are much less elevated in brain (x3) than in liver or spleen (x20) [62,64]. Myelin lipids are markedly affected in the NPC1 mouse model but in patients, a significant decrease is only seen in the early infantile neurological onset form [18,60].

#### ***Cell biology and cholesterol transport, and the brain enigma***

Initial studies by Peter Pentchev and associates and further work from several laboratories (reviewed in [10,20]) demonstrated, in cultured skin fibroblasts of Niemann-Pick C disease patients, a disruption in intracellular transport of endocytosed cholesterol. Endocytosed low density lipoproteins are delivered to late endosomes/lysosomes, where they are hydrolyzed, so that free cholesterol is released. In normal cells, this cholesterol is transported rapidly out of endosomes to the plasma membrane and the endoplasmic reticulum. In Niemann-Pick C disease cells (either NPC1 or NPC2), the cholesterol does not exit the endocytic pathway but accumulates within lysosomes. This anomaly constitutes the cellular hallmark of the disease. Due to this sequestration, the subsequent induction of all low-density lipoprotein cholesterol-mediated homeostatic responses (more specially cholesteryl ester formation) is retarded in Niemann-Pick C disease cells. Normal responses can be induced by membrane-permeable oxysterol and by mevalonate, showing that the ability of the cell to respond is maintained. Very recently, it was further shown that the block in cholesterol delivery to the ER can also be overcome by 2-hydroxypropyl-beta-cyclodextrin [65], and that this compound added to fibroblasts reduces the lysosomal cholesterol accumulation [66]. Studies in patients' cells

showed that lysosomal storage of unesterified cholesterol may show a variable intensity, and a “variant” biochemical phenotype with mild abnormalities has been described [67,68]. Later work showed that this phenotype was underlined by specific *NPC1* mutations (see below). Unexpectedly, fibroblasts from a large proportion of obligate heterozygotes have been found to show mild but definite changes [67-70].

This unique impairment in processing and utilization of endocytosed cholesterol obviously plays a key role in the pathogenesis of Niemann-Pick C disease, and, at least in extraneural organs, could actually explain a more general dysfunction of intracellular metabolism of lipids [63]. Sphingomyelin accumulation appears related to lysosomal cholesterol storage, since sphingomyelinase activity can be strongly modulated in fibroblast cultures of Niemann-Pick C disease patients by incubation in presence or absence of low-density lipoprotein. Cholesterol accumulation might also modulate glucosylceramide hydrolysis [71], as well as the trafficking of late endosomal proteins such as Rab 9 and mannose-6-phosphate receptors [72], two key players in the normal function of the endosomal/lysosomal system. There is thus good evidence that cholesterol accumulation in the late endosomal / lysosomal compartment can impair vesicular trafficking pathways.

The pathogenesis of the neuronal dysfunction appears by far more complex, since brain cholesterol is synthesized locally, mostly by oligodendroglial cells and to a lesser extent by astrocytes and neurons. Neurons might also acquire a small amount of cholesterol by glial delivery through apo-E uptake [73]. By chemical measurement, no significant increase of cholesterol concentrations can be found in dissected cerebral grey matter from human patients [60]. *In situ* labeling using filipin histochemistry, however, reveals a sequestration of unesterified cholesterol in cell bodies of neurons and glia of single NPC1 or NPC2 mutant mice as well as those of the double mutant [18, 58, 73-75]. These observations are not necessarily contradictory, since studies on cultured sympathetic neurons from NPC1 mutant mice gave indication that cholesterol did accumulate in cell bodies, but was decreased in distal axons, leading to a distribution imbalance [76,77]. One group has reported that endogenously synthesized cholesterol may significantly contribute to the overall cholesterol accumulation observed in Niemann-Pick C disease in various cell types, including glial cells [78]. Nevertheless, since abnormal filipin staining of neurons is also observed in a wide spectrum of other lysosomal storage disorders [discussed in 63], the exact participation of cellular cholesterol transport abnormalities in the pathophysiology of the neurodegenerative NP-C disease remains elusive. Of note, fibroblasts from patients with an adult-onset of neurological symptoms may show either a severe cholesterol trafficking defect or only minimal alterations (biochemical variant) [35,69,70]. Conversely, in two “variant” siblings who had died from a juvenile form, the liver showed no lipid accumulation (spleen did), but the brain showed typical accumulation of GM2 and GM3 gangliosides [79].

### ***The NPC1 and NPC2 proteins***

The mature native NPC1 is a large (1252 aminoacid) glycoprotein with 13 transmembrane domains, that resides primarily in late endosomes and interacts transiently with lysosomes and the trans-Golgi network [54,80]. It possesses a sterol-sensing domain (amino acid residues 615-797) showing homologies with those of HMG-CoA reductase, SCAP, patched and NPC1L, the exact role of which is still unclear although it appears necessary for protein function. Two luminal domains may play a role in protein-protein interactions: a cysteine-rich loop with a ring-finger motif which harbors about 1/3 of the mutations described in patients (amino acid residues 855-1098), and a highly conserved domain with a leucine-zipper motif, located in the N-terminal tail (amino acids 25-264) [81].

Importantly, the latter has been shown to possess a cholesterol-binding site (reviewed in [56]). Contrary to the NPC1 protein, the NPC2 protein is small (132 amino acids), soluble, secreted and recaptured. It is transported to the lysosome via the mannose-6-phosphate receptor and binds cholesterol with submicromolar affinity [56]. The mutation p.S120P (observed in a patient with a juvenile neurological onset and slowly progressive form [82]) has been instrumental to confirm the functional significance of the cholesterol-binding site of the NPC2 protein [83]. Studies in patients and animal models have shown that both NPC2 and NPC1 are required for cholesterol egress from the lysosome. Binding of cholesterol to NPC1 and dissociation both appear accelerated by NPC2 [83]. Based on the current stage of knowledge but fully compatible with earlier studies (reviewed in [56]) a “handoff” model has recently been proposed for the coordinated role of the two proteins [19]. In this model, cholesterol released within the lysosome binds to NPC2 with its hydroxyl group exposed; a transfer to the N-terminal domain of NPC1 occurs reversing its orientation, so that the hydrophobic side chain could lead the way into the membrane and/or the glycocalix. The most recent studies [65] indicate that the role of NPC2/NPC1 proteins in cholesterol transport is restricted to lysosomal export. Current data suggest that retrograde cholesterol movement from the plasma membrane to the ER does not require NPC1 [65] and implication of these proteins in cell processing of endogenously synthesized cholesterol [84] is still a matter of discussion.

Many uncertainties thus remain regarding the precise and complete functions of the NPC1 and NPC2 proteins. It has also been suggested that they could be involved in fusion/fission events between the late endosome and the lysosome. One important (and yet unanswered) question is whether these proteins – at least NPC1 - also directly regulate or mediate retrograde transport of other lysosomal cargo. Glycolipids, which constitute the main lipid accumulation in the brain, by opposition to the quantitatively minor cholesterol imbalance in neurons, are good candidates. The storage of GM2 and GM3 gangliosides in brain is not specific. Yet, the increase of GM2 occurs much earlier and is more prominent in NP-C than in other lysosomal diseases [63]. But no data supportive of a glycolipid transport by NPC1 or NPC2 have been published so far. It has also been postulated that sphingosine storage could be the primary trigger of a pathogenic cascade in NP-C since this lipid can disrupt calcium homeostasis in NPC1 lysosomes [85,86]. The latter studies, however, were conducted in non neural NP-C cells or in a drug (U18666A)-induced model. In brain, currently available data show a close link between accumulation of the different lipids, both in developmental terms and after therapeutic attempts [63,64]. No ganglioside or sphingosine accumulation can be detected in the brain of human fetuses at 24 gestational weeks, although the liver already shows a pronounced storage. Arguments for and against each of the accumulated lipids as the offending metabolite have recently been discussed [86]. Most likely, stored lipids (and possibly other metabolites) collectively contribute to the pathology. More work is clearly needed to better understand the cause of brain dysfunction in Niemann-Pick C disease. In particular, the mechanisms by which Purkinje cells and other neurons degenerate remain unclear.

### ***Disease-causing mutations and genotype-phenotype relationships***

The Niemann-Pick type C disease variation database [87] listed by January 2010 244 *NPC1* and 18 *NPC2* gene sequence variants. Reporting from diagnostic laboratories, however, has not been exhaustive, and the current number for identified *NPC1* disease-causing mutations is most likely close to 300. More than 60 polymorphisms of *NPC1* have also been described, some of them very common. In early genetic complementation studies, it was stated that about 95% of the families had mutations in the *NPC1* gene [14]. In France, among the 132 families genotyped so far, 9 had *NPC2* mutations. To date, only c:a 30 families have been

identified worldwide with mutations in the *NPC2* gene. Several large mutational studies have been published [33, 47,82,87-97], but only few functional studies [47,82,91,98-101].

The *NPC1* gene, mapped to chromosome 18q11-q12, spans 56 kb and contains 25 exons. One mutant allele, p.I1061T, is particularly frequent [26] (approximately 20-25 % of alleles in patients diagnosed in France or the United Kingdom). This mutation is also highly prevalent in patients from a Spanish-American isolate from the upper Rio Grande valley, but much less frequent in Portugal, Spain or Italy [91,94,96]. In the homoallelic state, it is associated with prominent cellular cholesterol trafficking abnormalities in fibroblasts of patients, and it correlates with a juvenile neurologic onset form of the disease [26]. In the heteroallelic state, so far it has never been found associated with the most severe infantile neurologic onset form. The I1061T mutant was shown to be a functional protein selected for endoplasmic reticulum-associated degradation due to protein misfolding and thus a potential target for chaperone therapy [98]. The second most recurrent *NPC1* mutation in Europe, p.P1007A, is the prototype of a “biochemical variant” mutation [47,89,95]. In the homozygous state, it has been described in a family with two adult onset siblings [91]. A number of other recurrent *NPC1* mutations seem to be associated with adult neurological onset of the disease [35,95]. The mutation p.G992W, typical of Nova-Scotian patients [17] is sporadically (but rarely) found in patients of other origin. As more patients are genotyped, a larger number of recurrent mutations are observed, some of them preferentially found in patients from defined ethnic origin.

The few genotype-phenotype studies published so far in NP-C1 patients generally showed good correlation between nonsense or frameshift mutations and the most severe neurologic course. Missense mutations have emphasized the functional significance of two particular domains of the NPC1 protein. Homozygous mutations in the sterol-sensing domain were found to be very deleterious, corresponding to a lack of mature NPC1 protein and to a very severe disease phenotype, biochemically and clinically [47]. The cysteine-rich luminal loop contains approximately one third of all described mutations, with a variable cellular and clinical impact [47,89,93,95]. Among others, it harbors the three most frequent mutations discussed above. Interestingly, mutations leading to a less severe impairment of cellular trafficking (“variant” phenotype) are typically located on this loop [47,90,91,93-95]. Genotype-phenotype correlations for more specific mutations have been discussed in earlier reports [11,88,95]

The *NPC2* gene (initially known as *HE1*), mapped to chromosome 14q24.3, spans 13.5 Kb and contains 5 exons [16]. One nonsense mutation (E20X) appears relatively frequent [82,92], and many other mutations also lead to a truncated protein. They have so far been associated with very severe clinical phenotypes. Described missense mutations have corresponded to more varied phenotypes, including juvenile and adult onset patients [82,92,93,101].

Finally, for both *NPC1* and *NPC2*, the study of a large number of multiplex families has clearly shown that mutations correlate with the neurological form of the disease, but not with the systemic manifestations [11].

## **Diagnostic methods**

The laboratory diagnostic algorithm proposed in a recent consensus report [27] is given in Fig. 3.

### ***Initial clinical assessment***

Suspecting Niemann-Pick disease type C is relatively easy in patients with the most typical symptoms, such as combined splenomegaly, ataxia, and supranuclear vertical gaze palsy. However, as described earlier, strikingly different clinical presentations exist, especially in infants and neonates. The fact that isolated spleno- or hepatosplenomegaly can be the presenting symptom long before neurologic onset has not been emphasized enough. Finally, the diagnosis is often very delayed (and probably often not made) in neurological cases lacking organomegaly, and in psychiatric cases. Consequently, the age at which the diagnosis is established is very variable. This is illustrated by data obtained in the author's laboratory for a representative cohort of patients (Fig. 4).

The characteristic key signs and symptoms in the systemic, ophthalmological and neuropsychiatric areas have been discussed above and the reader is also referred to a recent review [27]. A comprehensive clinical examination should be performed. The neurological evaluation must include muscle tone and strength tests, motor reflexes, assessment of movement (ataxia and dystonia) and swallowing testing [27]. Psychometric assessment is also important.

The ophthalmological assessment is of particular importance, because abnormal saccadic eye movements (SEM) are often the earliest neurological sign in NP-C. Proper examination is not always done, and reported findings are sometimes neglected in the global evaluation of the patient. The initial SEM deficit is in the vertical plane (downward, upward, or both). VSGP can be described as an increased latency of initiation of vertical saccades, with gradual slowing and eventual loss of saccadic velocity [27,102,103]. Subsequently, horizontal gaze is also affected. Cataplexy ranges from subtle signs (minor head-drop or falls, often confused with seizures) to full collapse in response to humorous stimuli [27].

### ***Neurophysiologic and neuroradiologic studies***

Hearing tests (audiogram and/or brainstem evoked potentials) often show abnormalities. MRI and CT scans are not very useful for diagnosis, as they may be normal or show cerebellar or cortical atrophy, or, in the severe infantile form, white matter changes. Some rare patients have been described with a peripheral neuropathy.

### ***Histology***

Foam cells and sea-blue histiocytes are usually – but not always - present in bone marrow. Foam cells stain strongly positive with filipin. Ultrastructural studies on skin [104], conjunctival, or liver biopsies can provide strong support for the diagnosis, but false-negative results often occur on liver biopsy studied by light microscopy only [41].

### ***Non-specific laboratory analyses***

Routine laboratory tests usually give normal results, except in patients with cholestatic jaundice or hypersplenism. Low HDL-cholesterol is a frequent but not universal finding. Plasma lipid profiles seem correlated to severity of cholesterol trafficking abnormalities [97]. Chitotriosidase activity is usually mildly elevated [105] but can be normal. Acid sphingomyelinase activity is normal or elevated in leukocytes (differential diagnosis with Niemann-Pick type B or atypical type A) but often partially deficient in fibroblasts [11,25,29,67].

### ***Specific laboratory diagnosis***

### *Biochemical/cell biology study: the “filipin” test*

The demonstration of impaired intracellular cholesterol transport and homeostasis is considered the primary diagnostic test for NP-C. These studies require living cells and thus a skin fibroblast culture. They should be conducted in specialized centres with the required experience. The “filipin test” is the most sensitive and specific assay. Fibroblasts are cultured in a LDL-enriched medium, then fixed and stained with filipin (a compound forming specific complexes with unesterified cholesterol). Fluorescence microscopic examination of NP-C-positive cells typically reveals numerous strongly fluorescent (cholesterol-filled) perinuclear vesicles. This “classical” storage pattern is observed in approximately 80-85% of cases. A lesser (and variable) level of storage is seen even under optimized conditions [67] in the remaining cases, described as having a “variant” biochemical phenotype [67,68]. As discussed above, several recurrent NPC1 mutations are known to result in this “variant” biochemical phenotype. Note that a similar, mildly abnormal filipin pattern, has been observed in a number of heterozygotes [69,70], but also not infrequently in acid sphingomyelinase deficiencies. Measurement of the LDL-induced rate of cholesteryl ester formation was until recently systematically used as a secondary test, showing very low levels in cell lines with a “classical” biochemical phenotype but only a mild or non-significant impairment in those with a “variant” phenotype [67,68]. As this test is complex, costly and time-consuming, mutation analysis is now often initiated directly when the filipin study is clearly positive. From the experience of the author, based on the study of cells from more than 600 NP-C patients, demonstration of cholesterol accumulation in cultured fibroblasts provides a clear-cut diagnosis in a majority of cases, but making a decision can be very difficult in some cell lines showing only minor abnormalities. In such cases, (and eventually in cases with apparently negative filipin but a history highly suggestive of NP-C), complementary mutation analysis is very useful to reach a definitive diagnosis.

### *Genetic testing*

It is highly advisable to undertake gene testing in every newly diagnosed patient, since molecular genetic study is today the highly preferred strategy for prenatal diagnosis, and the only reliable one for identification of carriers in blood relatives. Furthermore, as discussed above, gene testing can sometimes be necessary to confirm or disprove the diagnosis of NP-C. Genetic complementation studies - performed earlier in a few laboratories to define which gene was affected - are no longer used today, because cell hybridization and further testing are more elaborate than gene sequencing. Sequencing of all exons and boundaries is more laborious for the *NPC1* gene (25 exons) than for the *NPC2* gene (5 short exons), which is unfortunate, since over 95% of NP-C patients have pathological *NPC1* mutations. Rapid methods have been published to test for the two most frequent mutations [26,47]. Identification of *NPC1* mutations can, in some instances, be difficult and may require combined studies of gDNA and cDNA. All groups have met a common problem, namely that in some patients mutations could be identified in only one allele, and in a few of them, no mutation at all. The latter patients have raised the question of a potential third gene causing NP-C. This cannot be excluded, but often the possibilities of large deletions, or of deep intronic mutations [106] have not been investigated. Finally, due to the highly polymorphic nature of *NPC1*, interpretation of new missense mutations should be undertaken with caution.

### **Differential diagnosis**

In the neonate and young infant, Niemann-Pick disease type C must be differentiated from idiopathic neonatal hepatitis, and other causes of cholestatic icterus. Onset of cholestasis usually occurs in the early neonatal period. Associated splenomegaly is a useful orienting sign. In case of isolated splenomegaly or hepatosplenomegaly, NP-C should be considered as

a possible cause. Among other lipidoses, the most obvious differential diagnoses are Niemann-Pick type B (similar foam cells in bone marrow) and Gaucher disease. In older children and adults, depending on the symptoms, other conditions with cerebellar ataxia, dystonia, cataplexy and supranuclear gaze palsy need to be considered [27,31].

### **Genetic counseling**

Niemann-Pick C disease is genetically inherited following an autosomal recessive mode. The genetic status of a blood relative can be reliably established if mutations have been identified in the family index case. However, it is not currently possible to ascertain the status of a person from the general population, due to the complexity of *NPCI* gene sequencing and its polymorphic nature. Antenatal diagnosis is possible under the conditions described below.

The possibility of symptomatic heterozygotes has been raised in three families known to the author but ruled out in two of them (no further study in the third one). Two disease-causing *NPCI* mutations had been identified in each index case. In both families, the father of the proband developed progressive symptoms compatible with an adult onset neurologic form of NP-C. Subsequent complete gene sequencing revealed one allele carrying the mutation transmitted to the affected child, and another (not transmitted) disease-causing mutation on the other allele (M.T. Vanier and K. Harzer; M.T. Vanier and A. Ivanoiu, unpublished). These individuals were thus NP-C1 homozygotes with an adult onset form. These exceptional histories illustrate some of the problems eventually posed by the clinical heterogeneity of NP-C and the possible underestimation of adult-onset form of the disease.

### **Antenatal diagnosis**

Prenatal diagnosis of NP-C should be offered to couples at risk [27,107,108]. It is best achieved using chorionic villus sampling (CVS) at 10-12 weeks, but is also possible on amniotic cells. Molecular genetic analysis is today by far the preferred strategy [27], for several reasons. Unlike the cellular biology testing using filipin staining, it does not require cultured cells and a lengthy elaborate work up. The results can be obtained much earlier in pregnancy, and the tests can in principle be set up in any good molecular biology laboratory. It however requires that mutations have been identified on both alleles in the index case, or at least that suitable intragenic markers have been identified in the nuclear family. Today, few laboratories offer a prenatal test using the cellular biology strategy, which should be considered as a last resort due to its many drawbacks. Results will not be reached until 5-7 weeks after the sampling; the tests are technically difficult; besides, they are fully reliable only when the proband has shown severe abnormalities, thus excluding 15-20% of the families.

### **Management including treatment**

To date, management remains largely symptomatic. Information and support to families can be obtained through organizations specifically devoted to Niemann-Pick diseases (in the United States, United Kingdom, Germany, Spain, Italy, Argentina, Australia, Poland), to lysosomal diseases (France) or to inherited metabolic diseases (The Netherlands) [see appendix for websites]. Genetic counseling should be made available for family members. For detailed guidelines on current management of patients, the reader is referred to a recent publication compiled by an international working group [27]. A study on the cost of illness associated with NP-C in the UK has recently been published [109]

### ***Symptomatic management***

Seizures generally respond at least partially to antiepileptic drugs until a fairly advanced stage

of the disease. Cataplexy can usually be controlled by clomipramine, protriptyline, or modafinil. Anticholinergic agents have been reported to improve dystonia and tremor in some patients. Physiotherapy is useful in the management of spasticity and the prevention of contractures. Melatonin may be used to treat insomnia. Patients with a slow disease course may benefit from special schooling for handicapped children. Proper management of infections and of feeding difficulties (gastrostomy) is essential at an advanced stage of the disease.

### ***Specific treatment***

In the murine and feline NPC1 models, bone marrow transplantation (BMT) did not improve the neurological disease, not unexpectedly considering the properties of the NPC1 protein; similarly, after BMT the neurologic status of a child continued to deteriorate, although there was a regression of hepatosplenomegaly and lung infiltration [110]. In addition, liver transplantation performed in a few cases with cirrhosis did not influence the course of neurologic deterioration [111]. On the contrary, because the NPC2 protein is soluble, secreted and recaptured, there is a rationale supporting early hematopoietic stem cell transplantation in NP-C2 patients [82]. The long-term outcome is yet unknown, but encouraging results have recently been obtained in one patient transplanted at 18 months and followed up until 3 years of age [112].

Treatment strategies based on the hypothesis that cholesterol is the offending metabolite were first proposed in the early 90's. The combination of hypocholesterolemic drugs and a low-cholesterol diet seemed to partially reduce the cholesterol load in liver, but no amelioration of the neurological disease was seen in patients after 2 years of treatment [31].

Since glycolipid storage appears to contribute to at least some of the neuropathologic features, an iminosugar inhibitor of glucosylceramide synthase (miglustat, also known as N-butyl-deoxynojirimycin, NB-DNJ and OGT 918, later approved for substrate reduction therapy of mild to moderate type 1 Gaucher disease), was administered to *npc1* mutant mice and cats. It resulted in delayed onset of the neurological symptoms in both species, and a 20% longer survival of the mice [113]. A controlled clinical trial was thus initiated in neurologically symptomatic patients, first in adolescents and adults (12 years and above) [114], then in children (4-12 years). Long-term data from open-label extension treatment (up to 66 months) have now been reported in children [115] as well as in juvenile and adult patients [116] (reviewed in [27]). Overall, the disease course stabilized in 72% of patients treated for one year or more, based on a composite assessment of horizontal saccadic eye movement velocity, ambulation, swallowing and cognition. In January 2009, the European Union has extended the indication of miglustat to the treatment of progressive neurological manifestations in adult and pediatric patients with NP-C, and the drug is now approved for this indication in several other countries. This represents the first specific treatment for NP-C. Apart from single case reports [117,118], an international, multicenter observational cohort study in 66 patients treated off-label with miglustat has been published [119]. Evaluation made with a modified disease-specific disability scale [32] further showed a significant reduction in the annual rate of progression of the disease in a majority of patients. Late-onset forms generally appeared as the best responders. A further case series from Spain has been documented [120]. Longer term studies will be important to better evaluate the disease progression following the stabilisation phase [121]. Indication, clinical utility and monitoring of treatment with miglustat have been recently discussed [27,122]. In short, it has been recommended to treat patients as soon as they show neurological manifestations of any type. Due to the known adverse effects, such as diarrhea, flatulence, weight loss and tremor, it is not recommended

today to treat patients with systemic disease only. Note that miglustat is not expected to have an effect on the systemic manifestations of NP-C.

### ***Disease monitoring***

In order to monitor disease progression and, if applicable, patient responses to treatment, it is important to regularly quantify the degree of disability resulting from neurological impairment. Two disease-specific disability scales have recently been proposed [32,123]. The first one [32] (Table 1) evaluates four key parameters: ambulation, manipulation, language and swallowing, with a 4 to 5 point scale for each. This allows calculation of a composite score representing overall “functional disability”. Although not formally validated, it has already been used successfully in several cohort studies. Recent natural history surveys using these different scales both concluded to a linear clinical progression over time [123,124]. The cohort including a broader - and thus more representative - range of clinical phenotypes [124] showed a more rapid course in the patients with an early onset.

Useful monitoring tests have been recently discussed in detail [27]. Several methods for analysis of movement abnormalities [125,126] or neuropsychological profiles [127] have also been proposed. Results on three patients indicated that longitudinal MRS studies [128] might prove useful for follow up of therapy [129]. Diffusion tensor imaging has also been proposed [130].

### **Experimental therapeutic approaches in animal models**

Extensive research towards other therapeutic avenues is currently underway on animal and cellular models. These approaches have been reviewed in [27]. Various transgenic mice have been generated, such as mice over expressing Rab9, a protein involved in intracellular trafficking [131-133], or mice expressing NPC1 only in one particular brain cell type [134]. Most studies have however been conducted on the *npc1*<sup>nih</sup> mouse and a cat model (both spontaneous *npc1* mutants) [135,136], as well as a transgenic *npc2* mouse mutant [18]. These animals are particularly useful to study brain dysfunction, and facilitate various types of experiments, including administration of various compounds with a therapeutic goal. Data have been published in the mouse using imatinib [137], curcumin [85], non-steroid anti-inflammatory drugs [138], neurosteroids (allopregnanolone) in combination with 2-HP- $\beta$ -cyclodextrin [139], and with 2-HP- $\beta$ -cyclodextrin alone [64,140]. Chronic subcutaneous administration of high doses of 2-HP- $\beta$ -cyclodextrin resulted in a striking reduction of the various stored lipids both in the liver and the brain of NP-C mice, as well as a very significant effect on their lifespan [64]. An orphan drug designation has been sought for this compound from the US-FDA. However, translation of most of these studies to human patients is not straightforward. Even neglecting adverse effects [141] or the purity or homogeneity of certain compounds, a quite general and major limitation is the usual early timing of treatment (usually long before symptoms appear). Such experimental work in the whole animal is, however, important as it is generally felt that future treatment plans will combine several approaches and will be tailored to the individual.

### **Prognosis**

NP-C is a severe disorder that invariably leads to premature death, with few exceptions (three proven cases aged 53 years or more with isolated splenomegaly are known) [45-48]. However, as discussed above, the rate of progression and life span show considerable variation. The systemic disease can be fatal in early infancy. Patients with fetal hydrops survive at most a few days. Liver failure causes rapid death (before 3-6 months of age) in approximately 10% of neonates presenting with a cholestatic icterus, and a few patients (most

of them with a severe *NPC2* mutation) have died from severe pulmonary insufficiency. Neonatal cholestatic icterus is otherwise transient and usually resolves spontaneously by 4 months of age. Splenomegaly very rarely leads to hypersplenism. An important observation is that the age of onset of the systemic disease is generally unrelated to the subsequent neurological involvement and cannot be used as a predictor. This is well illustrated in Fig. 4, where several patients diagnosed in their first months of life are now teenagers. In the vast majority of patients, the lifespan is in large part determined by the age of onset of nervous system involvement. Data on large cohorts of patients recently compiled for Spain, the UK and France [27] are well in line with earlier reports. Patients with the severe neurologic early infantile form often die between 3 and 5 years of age, those with a late-infantile neurologic onset usually between 7 and 12. Patients with a juvenile neurologic onset survive until adolescence or later, with a sizable proportion reaching the age of 30. In a review of 68 cases with adult onset [35], the mean age at death (on 20 patients) was  $38 \pm 10.2$  years, but some patients have reached the age of 70. Motor involvement is often more severe and more rapidly progressive than mental retardation. Progressive and severe dysphagia requiring gastrostomy is a common complication. Severe and intractable epilepsy accelerates the downhill course of the disease. Psychiatric disturbances, in rare cases, may be prominent or even dramatic.

Regarding recurrence within a sibship, the study of many multiplex families has shown that as a rule, the neurological form - as defined by age of onset of neurological symptoms, and irrespective of the age of onset of the systemic disease - is similar between siblings. The subsequent course can however show variations, especially for cases developing severe epilepsy. On the other hand, there are many examples of families with one case of fetal hydrops or fatal neonatal liver disease and a sibling having a more classical neurovisceral form - more often of the early infantile type, but also of the late infantile or juvenile type.

Correlations between the neurological form and the severity of the cholesterol trafficking lesion as found by the filipin test has been discussed previously [11,67,70]. In brief, in the experience of the author, a "variant" biochemical phenotype tends to correlate with a less rapid course, since it has so far never been found in the most severe early infantile neurological form, is rare in late infantile forms, but seen in a number of juvenile and nearly half of the adult onset patients. On the other hand, finding a very severe cholesterol trafficking impairment (massive cholesterol accumulation in lysosomes) is not predictive of any form of the disease (seen in the other half of adult-onset patients).

Finally, although genotype-phenotype correlations are limited, in NP-C1, some degree of prediction is often possible. Thus far, the p.I1061T allele has not been associated with the most severe infantile neurological form [11,47]. Frameshift or nonsense mutations, as expected, but also missense mutations affecting the sterol sensing domain usually have a severe impact. On the other hand, association with a mutation leading (when in the homozygous state) to an adult onset form usually results in a slowly progressive juvenile or early adult onset form [95].

### **Unresolved questions**

NP-C is a disease with many unresolved questions. To begin with, the precise and complete function(s) of the NPC1 and NPC2 proteins are still largely unknown. Only few studies on cholesterol transport and metabolism have addressed the brain, in spite of the fact that brain has a cholesterol metabolism that is different from that in cells from systemic organs [142]. The nature of the primary offending metabolite in brain is also unknown. For these reasons, meaningful high throughput drug screening strategies are difficult to set up.

A major practical problem is the current lack of a biochemical test with sufficient specificity to be used for screening – or even better, diagnosis - that could be carried out on a blood sample. Having to start from a skin biopsy excludes NP-C from all “metabolic screens” and significantly contributes to the delay in diagnosis. Importantly, a recent pilot study indicates that plasma of patients with NP-C show a specific oxysterol profile that could be used as a biomarker [143]. This observation may impact the future diagnostic strategy.

As regards therapy, because the NPC1 protein, unlike many other lysosomal proteins, is not secreted and recaptured, many therapeutic strategies that are currently holding promises for the future seem not easily applicable to NP-C, including cell and gene therapy. Another difficulty to treat the brain dysfunction is the unknown nature of the primary targets. Along this line, the potential mode of action of some experimental compounds (among which is  $\beta$ -cyclodextrin) remains a puzzling question. Finally, the broad clinical spectrum, as well as the lack of good disease markers and clinical endpoints, makes evaluation of therapeutic trials particularly difficult.

## List of abbreviations

CT: computerized tomography; 2-HP- $\beta$ -cyclodextrin: 2-hydroxypropyl- $\beta$ -cyclodextrin; LDL: low-density lipoproteins; NP-C: Niemann-Pick type C; NP-C1: Niemann-Pick type C disease with mutations in the *NPC1* gene; NP-C2: Niemann-Pick type C disease with mutations in the *NPC2* gene; MRI: magnetic resonance imaging; MRS: magnetic resonance spectroscopy; VSGP: vertical supranuclear palsy.

## Competing interests

In the past 3 years, MTV has been an invited speaker in meetings organized and sponsored by Actelion, in postgraduate courses sponsored by Shire educational grants, and has served in an advisory board for Actelion. She has received occasional honoraria from Actelion and Shire.

## Appendix

Niemann-Pick diseases support groups and corresponding websites

### 1. Specific Niemann-Pick diseases support groups:

**UK:** Niemann-Pick disease Group (UK) [www.niemannpick.org.uk](http://www.niemannpick.org.uk)

**USA and Canada:** National Niemann-Pick disease Foundation (USA): [www.nnpdf.org](http://www.nnpdf.org) ; “Canadian chapter”: [www.nnpdf.ca](http://www.nnpdf.ca); Ara Parseghian Medical Research Foundation : [www.parseghian.org](http://www.parseghian.org)

**Germany:** Niemann-Pick Selbsthilfegruppe Deutschland : [www.niemann-pick.de](http://www.niemann-pick.de)

**Spain :** Fundacion Niemann-Pick de España : [www.fnp.es](http://www.fnp.es)

**Italy:** Associazione Italiana Niemann-Pick: [www.niemannpick.org](http://www.niemannpick.org)

**Australia:** Australian NPC disease Foundation: [www.npcd.org.au](http://www.npcd.org.au)

**Argentina:** Asociacion Niemann-Pick Argentina: [www.npc.org.ar](http://www.npc.org.ar)

**Poland :** Stowarzyszenie Chorych na NPC

*2. Support groups for Lysosomal Diseases or Inborn Errors of Metabolism with a specific Niemann-Pick subgroup:*

**France** (with antennas in French speaking areas of Belgium and Switzerland): Vaincre les Maladies Lysosomales [www.vml-asso.org](http://www.vml-asso.org)

**The Netherlands** :Volwassenen Kinderen en Stofwisselingsziekten : [www.stofwisselingsziekten.nl](http://www.stofwisselingsziekten.nl)

## **Acknowledgements**

Very special thanks are due to all members of the French “Comité pour le Traitement des Maladies de Niemann-Pick” (CETNP) and to Drs Gilles Millat and Philippe Latour. The author also wishes to acknowledge the large number of pediatricians, neuropediatricians and adult neurologists who have over the years provided clinical data on their patients. Finally, writing this review has been greatly facilitated by the knowledge gathered through longstanding and fruitful international collaborations with many colleagues. Work from the author’s laboratory discussed in this review has been mainly supported by INSERM, Vaincre les Maladies Lysosomales and an INSERM/AFM/French Ministry of Research grant (Research network on rare diseases, 4MR32F).

## References

1. Crocker AC, Farber S: **Niemann-Pick disease: a review of eighteen patients.** *Medicine (Baltimore)* 1958, **37**:1-95.
2. Crocker AC: **The cerebral defect in Tay-Sachs disease and Niemann-Pick disease.** *J Neurochem* 1961, **7**:69-80.
3. Brady RO, Kanfer JN, Mock MB, Fredrickson DS: **The metabolism of sphingomyelin. II. Evidence of an enzymatic deficiency in Niemann-Pick disease.** *Proc Natl Acad Sci U S A* 1966, **55**:366-369.
4. Pentchev PG, Boothe AD, Kruth HS, Weintraub H, Stivers J, Brady RO: **A genetic storage disorder in BALB/C mice with a metabolic block in esterification of exogenous cholesterol.** *J Biol Chem* 1984, **259**:5784-5791.
5. Pentchev PG, Brady RO, Blanchette-Mackie EJ, Vanier MT, Carstea ED, Parker CC, Goldin E, Roff CF: **The Niemann-Pick C lesion and its relationship to the intracellular distribution and utilization of LDL cholesterol.** *Biochim Biophys Acta* 1994, **1225**:235-243.
6. Pentchev PG, Comly ME, Kruth HS, Vanier MT, Wenger DA, Patel S, Brady RO: **A defect in cholesterol esterification in Niemann-Pick disease (type C) patients.** *Proc Natl Acad Sci U S A* 1985, **82**:8247-8251.
7. Pentchev PG, Comly ME, Kruth HS, Tokoro T, Butler J, Sokol J, Filling-Katz M, Quirk JM, Marshall DC, Patel S, Vanier MT, Brady RO: **Group C Niemann-Pick disease: faulty regulation of low-density lipoprotein uptake and cholesterol storage in cultured fibroblasts.** *FASEB J* 1987, **1**:40-45.
8. Sokol J, Blanchette-Mackie J, Kruth HS, Dwyer NK, Amende LM, Butler JD, Robinson E, Patel S, Brady RO, Comly ME, Vanier MT, Pentchev PG: **Type C Niemann-Pick disease. Lysosomal accumulation and defective intracellular mobilization of low density lipoprotein cholesterol.** *J Biol Chem* 1988, **263**:3411-3417.
9. Liscum L, Ruggiero RM, Faust JR: **The intracellular transport of low density lipoprotein-derived cholesterol is defective in Niemann-Pick type C fibroblasts.** *J Cell Biol* 1989, **108**:1625-1636.
10. Patterson MC, Vanier MT, Suzuki K, Morris JA, Carstea E, Neufeld EB, Blanchette-Mackie JE, Pentchev P: **Niemann-Pick disease type C: a lipid trafficking disorder.** In *The Metabolic and Molecular Bases of Inherited Disease*, 8<sup>th</sup> Edition. Edited by Scriver CR, Beaudet AL, Sly WS, Valle D, Childs B, Kinzler KW, Vogelstein B. New York: Mc Graw Hill; 2001:3611-3634.
11. Vanier MT, Millat G: **Niemann-Pick disease type C.** *Clin Genet* 2003, **64**:269-281.
12. Ioannou YA: **Guilty until proven innocent: the case of NPC1 and cholesterol.** *Trends Biochem Sci* 2005, **30**:498-505.
13. Steinberg SJ, Ward CP, Fensom AH: **Complementation studies in Niemann-Pick disease type C indicate the existence of a second group.** *J Med Genet* 1994, **31**:317-320.
14. Vanier MT, Duthel S, Rodriguez-Lafrasse C, Pentchev P, Carstea ED: **Genetic heterogeneity in Niemann-Pick C disease: a study using somatic cell hybridization and linkage analysis.** *Am J Hum Genet* 1996, **58**:118-125.
15. Carstea ED, Morris JA, Coleman KG, Loftus SK, Zhang D, Cummings C, Gu J, Rosenfeld MA, Pavan WJ, Krizman DB, Nagle J, Polymeropoulos MH, Sturley SL, Ioannou YA, Higgins ME, Comly M, Cooney A, Brown A, Kaneski CR, Blanchette-Mackie EJ, Dwyer NK, Neufeld EB, Chang TY, Liscum L, Strauss JF, III, Ohno K, Zeigler M, Carmi R, Sokol J, Markie D, O'Neill RR, van Diggelen OP, Elleder M, Patterson MC, Brady RO, Vanier MT, Pentchev PG, Tagle DA: **Niemann-Pick C1 disease gene: homology to mediators of cholesterol homeostasis.** *Science* 1997, **277**:228-231.
16. Naureckiene S, Sleat DE, Lackland H, Fensom A, Vanier MT, Wattiaux R, Jadot M, Lobel P: **Identification of HE1 as the second gene of Niemann-Pick C disease.** *Science* 2000, **290**:2298-2301.
17. Greer WL, Riddell DC, Gillan TL, Girouard GS, Sparrow SM, Byers DM, Dobson MJ, Neumann PE: **The Nova Scotia (type D) form of Niemann-Pick disease is caused by a G3097-->T transversion in NPC1.** *Am J Hum Genet* 1998, **63**:52-54.
18. Sleat DE, Wiseman JA, El Banna M, Price SM, Verot L, Shen MM, Tint GS, Vanier MT, Walkley SU, Lobel P: **Genetic evidence for nonredundant functional cooperativity between NPC1 and NPC2 in lipid transport.** *Proc Natl Acad Sci U S A* 2004, **101**:5886-5891.
19. Kwon HJ, Abi-Mosleh L, Wang ML, Deisenhofer J, Goldstein JL, Brown MS, Infante RE: **Structure of N-terminal domain of NPC1 reveals distinct subdomains for binding and transfer of cholesterol.** *Cell* 2009, **137**:1213-1224.
20. Pentchev PG, Vanier MT, Suzuki K, Patterson M: **Niemann-Pick disease type C: a cellular cholesterol lipidosis.** In *The Metabolic and Molecular Bases of Inherited Disease*, 7<sup>th</sup> edition. Edited by Scriver CR, Beaudet AL, Sly WS, Valle D. New York: McGraw Hill; 1995: 2625-2639.
21. Meikle PJ, Hopwood JJ, Clague AE, Carey WF: **Prevalence of lysosomal storage disorders.** *JAMA* 1999, **281**:249-254.

22. Poorthuis BJ, Wevers RA, Kleijer WJ, Groener JE, de Jong JG, van Weely S, Niezen-Koning KE, van Diggelen OP: **The frequency of lysosomal storage diseases in The Netherlands.** *Hum Genet* 1999, **105**:151-156.
23. Pinto R, Caseiro C, Lemos M, Lopes L, Fontes A, Ribeiro H, Pinto E, Silva E, Rocha S, Marcao A, Ribeiro I, Lacerda L, Ribeiro G, Amaral O, Sa Miranda MC: **Prevalence of lysosomal storage diseases in Portugal.** *Eur J Hum Genet* 2004, **12**:87-92.
24. Winsor EJ, Welch JP: **Genetic and demographic aspects of Nova Scotia Niemann-Pick disease (type D).** *Am J Hum Genet* **30**: 530-538.
25. Wenger DA, Barth G, Githens JH: **Nine cases of sphingomyelin lipidosis, a new variant in Spanish-American Children. Juvenile variant of Niemann-Pick Disease with foamy and sea-blue histiocytes.** *Am J Dis Child* 1977, **131**:955-961.
26. Millat G, Marçais C, Rafi MA, Yamamoto T, Morris JA, Pentchev PG, Ohno K, Wenger DA, Vanier MT: **Niemann-Pick C1 disease: the I1061T substitution is a frequent mutant allele in patients of Western European descent and correlates with a classic juvenile phenotype.** *Am J Hum Genet* 1999, **65**:1321-1329.
27. Wraith JE, Baumgartner MR, Bembi B, Covanis A, Levade T, Mengel E, Pineda M, Sedel F, Topcu M, Vanier MT, Widner H, Wijburg FA, Patterson MC: **Recommendations on the diagnosis and management of Niemann-Pick disease type C.** *Mol Genet Metab* 2009, **98**:152-165.
28. Spiegel R, Raas-Rothschild A, Reish O, Regev M, Meiner V, Bargal R, Sury V, Meir K, Nadjari M, Hermann G, Iancu TC, Shalev SA, Zeigler M: **The clinical spectrum of fetal Niemann-Pick type C.** *Am J Med Genet A* 2009, **149A**:446-450.
29. Vanier MT, Wenger DA, Comly ME, Rousson R, Brady RO, Pentchev PG: **Niemann-Pick disease group C: clinical variability and diagnosis based on defective cholesterol esterification. A collaborative study on 70 patients.** *Clin Genet* 1988, **33**:331-348.
30. Trendelenburg G, Vanier MT, Maza S, Millat G, Bohner G, Munz DL, Zschenderlein R: **Niemann-Pick type C disease in a 68-year-old patient.** *J Neurol Neurosurg Psychiatry* 2006, **77**:997-998.
31. Schiffmann R: **Niemann-Pick disease type C. From bench to bedside.** *JAMA* 1996, **276**:561-564.
32. Iturriaga C, Pineda M, Fernandez-Valero EM, Vanier MT, Coll MJ: **Niemann-Pick C disease in Spain: clinical spectrum and development of a disability scale.** *J Neurol Sci* 2006, **249**:1-6.
33. Imrie J, Dasgupta S, Besley GT, Harris C, Heptinstall L, Knight S, Vanier MT, Fensom AH, Ward C, Jacklin E, Whitehouse C, Wraith JE: **The natural history of Niemann-Pick disease type C in the UK.** *J Inherit Metab Dis* 2007, **30**:51-59.
34. Garver WS, Francis GA, Jelinek D, Shepherd G, Flynn J, Castro G, Walsh VC, Coppock DL, Pettit KM, Heidenreich RA, Meaney FJ: **The National Niemann-Pick C1 disease database: report of clinical features and health problems.** *Am J Med Genet A* 2007, **143A**:1204-1211.
35. Sevin M, Lesca G, Baumann N, Millat G, Lyon-Caen O, Vanier MT, Sedel F: **The adult form of Niemann-Pick disease type C.** *Brain* 2007, **130**:120-133.
36. Solomon D, Winkelman AC, Zee DS, Gray L, Buttner-Ennever J: **Niemann-Pick type C disease in two affected sisters: ocular motor recordings and brain-stem neuropathology.** *Ann N Y Acad Sci* 2005, **1039**:436-445.
37. Kandt RS, Emerson RG, Singer HS, Valle DL, Moser HW: **Cataplexy in variant forms of Niemann-Pick disease.** *Ann Neurol* 1982, **12**:284-288.
38. Oyama K, Takahashi T, Shoji Y, Oyamada M, Noguchi A, Tamura H, Takada G, Kanbayashi T: **Niemann-Pick disease type C: cataplexy and hypocretin in cerebrospinal fluid.** *Tohoku J Exp Med* 2006, **209**:263-267.
39. Imrie J, Wraith JE: **Isolated splenomegaly as the presenting feature of Niemann-Pick disease type C.** *Arch Dis Child* 2001, **84**:427-429.
40. Vanier MT, Suzuki K: **Niemann-Pick diseases.** In: *Neurodystrophies and Neurolipidoses* Edited by Moser HW. Handbook of Clinical Neurology. Vol. 66/Revised Series Vol. 22. Amsterdam: Elsevier Science; 1996:133-162.
41. Kelly DA, Portmann B, Mowat AP, Sherlock S, Lake BD: **Niemann-Pick disease type C: diagnosis and outcome in children, with particular reference to liver disease.** *J Pediatr* 1993, **123**:242-247.
42. Yerushalmi B, Sokol RJ, Narkewicz MR, Smith D, Ashmead JW, Wenger DA: **Niemann-Pick disease type C in neonatal cholestasis at a North American Center.** *J Pediatr Gastroenterol Nutr* 2002, **35**:44-50.
43. Bjurulf B, Spletalen S, Erichsen A, Vanier MT, Strom EH, Stromme P: **Niemann-Pick disease type C2 presenting as fatal pulmonary alveolar lipoproteinosis: morphological findings in lung and nervous tissue.** *Med Sci Monit* 2008, **14**:CS71-CS75.44.
44. Griese M, Brasch F, Aldana VR, Cabrera MM, Goelnitz U, Ikonen E, Karam BJ, Liebisch G, Linder MD, Lohse P, Meyer W, Schmitz G, Pamir A, Ripper J, Rolfs A, Schams A, Lezana FJ: **Respiratory disease in Niemann-Pick type C2 is caused by pulmonary alveolar proteinosis.** *Clin Genet* 2010, **77**:119-130.

45. Fensom AH, Grant AR, Steinberg SJ, Ward CP, Lake BD, Logan EC, Hulman G: **An adult with a non-neuronopathic form of Niemann-Pick C disease.** *J Inherit Metab Dis* 1999, **22**:84-86.
46. Fröhlich E, Harzer K, Heller T, Ruhl U: **Sonographisch echodichte Milztumoren: Knotige Manifestation eines Morbus Niemann-Pick Typ C [Ultrasound echogenic splenic tumors: nodular manifestation of type C Niemann-Pick disease].** *Ultraschall Med* 1990, **11**:119-122.
47. Millat G, Marçais C, Tomasetto C, Chikh K, Fensom AH, Harzer K, Wenger DA, Ohno K, Vanier MT: **Niemann-Pick C1 disease: correlations between NPC1 mutations, levels of NPC1 protein, and phenotypes emphasize the functional significance of the putative sterol-sensing domain and of the cysteine-rich luminal loop.** *Am J Hum Genet* 2001, **68**:1373-1385.
48. Dvorakova L, Sikora J, Hrebicek M, Hulkova H, Bouckova M, Stolnaja L, Elleder M: **Subclinical course of adult visceral Niemann-Pick type C1 disease. A rare or underdiagnosed disorder?** *J Inherit Metab Dis* 2006, **29**:591.
49. Shulman LM, David NJ, Weiner WJ: **Psychosis as the initial manifestation of adult-onset Niemann-Pick disease type C.** *Neurology* 1995, **45**:1739-1743.
50. Imrie J, Vijayaraghaven S, Whitehouse C, Harris S, Heptinstall L, Church H, Cooper A, Besley GT, Wraith JE: **Niemann-Pick disease type C in adults.** *J Inherit Metab Dis* 2002, **25**:491-500.
51. Klünemann HH, Elleder M, Kaminski WE, Snow K, Peyser JM, O'Brien JF, Munoz D, Schmitz G, Klein HE, Pendlebury WW: **Frontal lobe atrophy due to a mutation in the cholesterol binding protein HE1/NPC2.** *Ann Neurol* 2002, **52**:743-749.
52. Walterfang M, Fietz M, Fahey M, Sullivan D, Leane P, Lubman DI, Velakoulis D: **The neuropsychiatry of Niemann-Pick type C disease in adulthood.** *J Neuropsychiatry Clin Neurosci* 2006, **18**:158-170.
53. Klarner B, Klünemann HH, Lurding R, Aslanidis C, Rupprecht R: **Neuropsychological profile of adult patients with Niemann-Pick C1 (NPC1) mutations.** *J Inherit Metab Dis* 2007, **30**:60-67.
54. Higgins ME, Davies JP, Chen FW, Ioannou YA: **Niemann-Pick C1 is a late endosome-resident protein that transiently associates with lysosomes and the trans-Golgi network.** *Mol Genet Metab* 1999, **68**:1-13.
55. Vanier MT, Millat G: **Structure and function of the NPC2 protein.** *Biochim Biophys Acta* 2004, **1685**:14-21.
56. Storch J, Xu Z: **Niemann-Pick C2 (NPC2) and intracellular cholesterol trafficking.** *Biochim Biophys Acta* 2009, **1791**:671-678.
57. Vincent I, Bu B, Erickson RP: **Understanding Niemann-Pick type C disease: a fat problem.** *Curr Opin Neurol* 2003, **16**:155-161.
58. Walkley SU, Suzuki K: **Consequences of NPC1 and NPC2 loss of function in mammalian neurons.** *Biochim Biophys Acta* 2004, **1685**:48-62.
59. Vanier MT: **Biochemical studies in Niemann-Pick disease. I. Major sphingolipids of liver and spleen.** *Biochim Biophys Acta* 1983, **750**:178-184.
60. Vanier MT: **Lipid changes in Niemann-Pick disease type C brain: personal experience and review of the literature.** *Neurochem Res* 1999, **24**:481-489.
61. Goldin E, Roff CF, Miller SP, Rodriguez-Lafrasse C, Vanier MT, Brady RO, Pentchev PG: **Type C Niemann-Pick disease: a murine model of the lysosomal cholesterol lipidosis accumulates sphingosine and sphinganine in liver.** *Biochim Biophys Acta* 1992, **1127**:303-311.
62. Rodriguez-Lafrasse C, Rousson R, Pentchev PG, Louisot P, Vanier MT: **Free sphingoid bases in tissues from patients with type C Niemann-Pick disease and other lysosomal storage disorders.** *Biochim Biophys Acta* 1994, **1226**:138-144.
63. Walkley SU, Vanier MT: **Secondary lipid accumulation in lysosomal disease.** *Biochim Biophys Acta* 2009, **1793**:726-736.
64. Davidson CD, Ali NF, Micsenyi MC, Stephney G, Renault S, Dobrenis K, Ory DS, Vanier MT, Walkley SU: **Chronic cyclodextrin treatment of murine Niemann-Pick C disease ameliorates neuronal cholesterol and glycosphingolipid storage and disease progression.** *PLoS One* 2009, **4**:e6951.
65. Abi-Mosleh L, Infante RE, Radhakrishnan A, Goldstein JL, Brown MS: **Cyclodextrin overcomes deficient lysosome-to-endoplasmic reticulum transport of cholesterol in Niemann-Pick type C cells.** *Proc Natl Acad Sci U S A* 2009, **106**:19316-19321.
66. Rosenbaum AI, Zhang G, Warren JD, Maxfield FR: **Endocytosis of beta-cyclodextrins is responsible for cholesterol reduction in Niemann-Pick type C mutant cells.** *Proc Natl Acad Sci U S A* 2010, **107**:5477-5482.
67. Vanier MT, Rodriguez-Lafrasse C, Rousson R, Gazzah N, Juge MC, Pentchev PG, Revol A, Louisot P: **Type C Niemann-Pick disease: spectrum of phenotypic variation in disruption of intracellular LDL-derived cholesterol processing.** *Biochim Biophys Acta* 1991, **1096**:328-337.
68. Argoff CE, Comly ME, Blanchette-Mackie J, Kruth HS, Pye HT, Goldin E, Kaneski C, Vanier MT, Brady RO, Pentchev PG: **Type C Niemann-Pick disease: cellular uncoupling of cholesterol homeostasis is**

- linked to the severity of disruption in the intracellular transport of exogenously derived cholesterol.** *Biochim Biophys Acta* 1991, **1096**:319-327.
69. Vanier MT: **Phenotypic and genetic heterogeneity in Niemann-Pick disease type C: current knowledge and practical implications** . *Wien Klin Wochenschr* 1997, **109**:68-73.
  70. Vanier MT, Suzuki K: **Recent advances in elucidating Niemann-Pick C disease.** *Brain Pathol* 1998, **8**:163-174
  71. Salvioli R, Scarpa S, Ciaffoni F, Tatti M, Ramoni C, Vanier MT, Vaccaro AM: **Glucosylceramidase mass and subcellular localization are modulated by cholesterol in Niemann-Pick disease type C.** *J Biol Chem* 2004, **279**:17674-17680.
  72. Ganley IG, Pfeffer SR: **Cholesterol accumulation sequesters Rab9 and disrupts late endosome function in NPC1-deficient cells.** *J Biol Chem* 2006, **281**:17890-17899.
  73. Vance JE, Hayashi H, Karten B: **Cholesterol homeostasis in neurons and glial cells.** *Semin Cell Dev Biol* 2005, **16**:193-212.
  74. Reid PC, Sakashita N, Sugii S, Ohno-Iwashita Y, Shimada Y, Hickey WF, Chang TY: **A novel cholesterol stain reveals early neuronal cholesterol accumulation in the Niemann-Pick type C1 mouse brain.** *J Lipid Res* 2004, **45**:582-591.
  75. Liu Y, Wu YP, Wada R, Neufeld EB, Mullin KA, Howard AC, Pentchev PG, Vanier MT, Suzuki K, Proia RL: **Alleviation of neuronal ganglioside storage does not improve the clinical course of the Niemann-Pick C disease mouse.** *Hum Mol Genet* 2000, **9**:1087-1092.
  76. Karten B, Vance DE, Campenot RB, Vance JE: **Cholesterol accumulates in cell bodies, but is decreased in distal axons, of Niemann-Pick C1-deficient neurons.** *J Neurochem* 2002, **83**:1154-1163.
  77. Karten B, Peake KB, Vance JE: **Mechanisms and consequences of impaired lipid trafficking in Niemann-Pick type C1-deficient mammalian cells.** *Biochim Biophys Acta* 2009, **1791**:659-670.
  78. Reid PC, Sugii S, Chang TY: **Trafficking defects in endogenously synthesized cholesterol in fibroblasts, macrophages, hepatocytes, and glial cells from Niemann-Pick type C1 mice.** *J Lipid Res* 2003, **44**:1010-9.
  79. Martin JJ, Lowenthal A, Ceuterick C, Vanier MT: **Juvenile dystonic lipidosis (variant of Niemann-Pick disease type C)** . *J Neurol Sci* 1984, **66**:33-45.
  80. Neufeld EB, Wastney M, Patel S, Suresh S, Cooney AM, Dwyer NK, Roff CF, Ohno K, Morris JA, Carstea ED, Incardona JP, Strauss JF, III, Vanier MT, Patterson MC, Brady RO, Pentchev PG, Blanchette-Mackie EJ: **The Niemann-Pick C1 protein resides in a vesicular compartment linked to retrograde transport of multiple lysosomal cargo.** *J Biol Chem* 1999, **274**:9627-9635
  81. Davies JP, Ioannou YA: **Topological analysis of Niemann-Pick C1 protein reveals that the membrane orientation of the putative sterol-sensing domain is identical to those of 3-hydroxy-3-methylglutaryl-CoA reductase and sterol regulatory element binding protein cleavage-activating protein.** *J Biol Chem* 2000, **275**:24367-24374
  82. Verot L, Chikh K, Freydiere E, Honore R, Vanier MT, Millat G: **Niemann-Pick C disease: functional characterization of three NPC2 mutations and clinical and molecular update on patients with NPC2.** *Clin Genet* 2007, **71**:320-330.
  83. Infante RE, Wang ML, Radhakrishnan A, Kwon HJ, Brown MS, Goldstein JL: **NPC2 facilitates bidirectional transfer of cholesterol between NPC1 and lipid bilayers, a step in cholesterol egress from lysosomes.** *Proc Natl Acad Sci U S A* 2008, **105**:15287-15292.
  84. Cruz JC, Chang TY: **Fate of endogenously synthesized cholesterol in Niemann-Pick type C1 cells.** *J Biol Chem* 2000, **275**:41309-41316
  85. Lloyd-Evans E, Morgan AJ, He X, Smith DA, Elliot-Smith E, Sillence DJ, Churchill GC, Schuchman EH, Galione A, Platt FM: **Niemann-Pick disease type C1 is a sphingosine storage disease that causes deregulation of lysosomal calcium.** *Nat Med* 2008, **14**:1247-1255.
  86. Lloyd-Evans E, Platt FM: **Lipids on Trial: The Search for the Offending Metabolite in Niemann-Pick type C Disease.** *Traffic* 2010, **11**:419-428.
  87. Runz H, Dolle D, Schlitter AM, Zschocke J: **NPC-db, a Niemann-Pick type C disease gene variation database.** *Hum Mutat* 2008, **29**:345-350. <http://npc.fzk.de>
  88. Yamamoto T, Nanba E, Ninomiya H, Higaki K, Taniguchi M, Zhang H, Akaboshi S, Watanabe Y, Takeshima T, Inui K, Okada S, Tanaka A, Sakuragawa N, Millat G, Vanier MT, Morris JA, Pentchev PG, Ohno K: **NPC1 gene mutations in Japanese patients with Niemann-Pick disease type C.** *Hum Genet* 1999, **105**:1016.
  89. Greer WL, Dobson MJ, Girouard GS, Byers DM, Riddell DC, Neumann PE: **Mutations in NPC1 highlight a conserved NPC1-specific cysteine-rich domain.** *Am J Hum Genet* 1999, **65**:1252-1260.
  90. Sun X, Marks DL, Park WD, Wheatley CL, Puri V, O'Brien JF, Kraft DL, Lundquist PA, Patterson MC, Pagano RE, Snow K: **Niemann-Pick C variant detection by altered sphingolipid trafficking and correlation with mutations within a specific domain of NPC1.** *Am J Hum Genet* 2001, **68**:1361-1372.

91. Ribeiro I, Marcao A, Amaral O, Sa Miranda MC, Vanier MT, Millat G: **Niemann-Pick type C disease: NPC1 mutations associated with severe and mild cellular cholesterol trafficking alterations.** *Hum Genet* 2001, **109**:24-32.
92. Millat G, Chikh K, Naureckiene S, Sleat DE, Fensom AH, Higaki K, Elleder M, Lobel P, Vanier MT: **Niemann-Pick disease type C: spectrum of HE1 mutations and genotype/phenotype correlations in the NPC2 group.** *Am J Hum Genet* 2001, **69**:1013-1021.
93. Park WD, O'Brien JF, Lundquist PA, Kraft DL, Vockley CW, Karnes PS, Patterson MC, Snow K: **Identification of 58 novel mutations in Niemann-Pick disease type C: correlation with biochemical phenotype and importance of PTC1-like domains in NPC1.** *Hum Mutat* 2003, **22**:313-325.
94. Fernandez-Valero EM, Ballart A, Iturriaga C, Lluch M, Macias J, Vanier MT, Pineda M, Coll MJ: **Identification of 25 new mutations in 40 unrelated Spanish Niemann-Pick type C patients: genotype-phenotype correlations.** *Clin Genet* 2005, **68**:245-254.
95. Millat G, Bailo N, Molinero S, Rodriguez C, Chikh K, Vanier MT: **Niemann-Pick C disease: use of denaturing high performance liquid chromatography for the detection of NPC1 and NPC2 genetic variations and impact on management of patients and families.** *Mol Genet Metab* 2005, **86**:220-232.
96. Fancello T, Dardis A, Rosano C, Tarugi P, Tappino B, Zampieri S, Pinotti E, Corsolini F, Fecarotta S, D'Amico A, Di Rocco M, Uziel G, Calandra S, Bembi B, Filocamo M: **Molecular analysis of NPC1 and NPC2 gene in 34 Niemann-Pick C Italian patients: identification and structural modeling of novel mutations.** *Neurogenetics* 2009, **10**:229-239.
97. Garver WS, Jelinek D, Meaney FJ, Flynn J, Pettit KM, Shepard G, Heidenreich RA, Walsh Vockley CM, Castro G, Francis GA: **The National Niemann-Pick Type C1 disease database: Correlation of lipid profiles, mutations, and biochemical phenotypes.** *J Lipid Res* 2009.
98. Gelsthorpe ME, Baumann N, Millard E, Gale SE, Langmade SJ, Schaffer JE, Ory DS: **Niemann-Pick type C1 I1061T mutant encodes a functional protein that is selected for endoplasmic reticulum-associated degradation due to protein misfolding.** *J Biol Chem* 2008, **283**:8229-8236.
99. Blom TS, Linder MD, Snow K, Pihko H, Hess MW, Jokitalo E, Veckman V, Syvanen AC, Ikonen E: **Defective endocytic trafficking of NPC1 and NPC2 underlying infantile Niemann-Pick type C disease.** *Hum Mol Genet* 2003, **12**:257-272.
100. Macias-Vidal J, Gort L, Lluch M, Pineda M, Coll MJ: **Nonsense-mediated mRNA decay process in nine alleles of Niemann-Pick type C patients from Spain.** *Mol Genet Metab* 2009, **97**:60-64..
101. Chikh K, Rodriguez C, Vey S, Vanier MT, Millat G: **Niemann-Pick type C disease: subcellular location and functional characterization of NPC2 proteins with naturally occurring missense mutations.** *Hum Mutat* 2005, **26**:20-28.
102. Rottach KG, von Maydell RD, Das VE, Zivotofsky AZ, Discenna AO, Gordon JL, Landis DM, Leigh RJ: **Evidence for independent feedback control of horizontal and vertical saccades from Niemann-Pick type C disease.** *Vision Res* 1997, **37**:3627-3638.
103. Abel LA, Walterfang M, Fietz M, Bowman EA, Velakoulis D: **Saccades in adult Niemann-Pick disease type C reflect frontal, brainstem, and biochemical deficits.** *Neurology* 2009, **72**:1083-1086.
104. Boustany RN, Kaye E, Alroy J: **Ultrastructural findings in skin from patients with Niemann-Pick disease, type C.** *Pediatr Neurol* 1990, **6**:177-183.
105. Ries M, Schaefer E, Luhrs T, Mani L, Kuhn J, Vanier MT, Krummenauer F, Gal A, Beck M, Mengel E: **Critical assessment of chitotriosidase analysis in the rational laboratory diagnosis of children with Gaucher disease and Niemann-Pick disease.** *J Inher Metab Dis* 2006, **29** : 647-652.
106. Rodriguez-Pascau L, Coll MJ, Vilageliu L, Grinberg D: **Antisense oligonucleotide treatment for a pseudoexon-generating mutation in the NPC1 gene causing Niemann-Pick type C diseaseb.** *Hum Mutat* 2009,**30**:1117-1122.
107. Vanier MT, Rodriguez-Lafrasse C, Rousson R, Mandon G, Boue J, Choiset A, Peyrat MF, Dumontel C, Juge MC, Pentchev PG, Revol A, Louisot P. **Prenatal diagnosis of Niemann-Pick type C disease: current strategy from an experience of 37 pregnancies at risk.** *Am J Hum Genet* 1992, **51**:111-122.
108. Vanier MT: **Prenatal diagnosis of Niemann-Pick diseases types A, B and C.** *Prenat Diagn* 2002, **22**:630-632.
109. Imrie J, Galani C, Gairy K, Lock K, Hunsche E: **Cost of illness associated with Niemann-Pick disease type C in the UK.** *J Med Econ* 2009, **12**:219-229.
110. Hsu YS, Hwu WL, Huang SF, Lu MY, Chen RL, Lin DT, Peng SS, Lin KH: **Niemann-Pick disease type C (a cellular cholesterol lipidosis) treated by bone marrow transplantation.** *Bone Marrow Transplant* 1999, **24**:103-107.
111. Gartner JC, Jr., Bergman I, Malatack JJ, Zitelli BJ, Jaffe R, Watkins JB, Shaw BW, Iwatsuki S, Starzl TE: **Progression of neurovisceral storage disease with supranuclear ophthalmoplegia following orthotopic liver transplantation.** *Pediatrics* 1986, **77**:104-106.

112. Bonney DK, O'Meara A, Shabani A, Imrie J, Bigger BW, Jones S, Wraith JE, Wynn RF: **Successful allogeneic bone marrow transplant for Niemann-Pick disease type C2 is likely to be associated with a severe "graft versus substrate" effect.** *J Inherit Metab Dis* 2010, e-pub ahead of press
113. Zervas M, Somers KL, Thrall MA, Walkley SU: **Critical role for glycosphingolipids in Niemann-Pick disease type C.** *Curr Biol* 2001, **11**:1283-1287.
114. Patterson MC, Vecchio D, Prady H, Abel L, Wraith JE: **Miglustat for treatment of Niemann-Pick C disease: a randomised controlled study.** *Lancet Neurol* 2007, **6**:765-772.
115. Patterson MC, Vecchio D, Jacklin E, Abel L, Chadha-Boreham H, Luzy C, Giorgino R, Wraith JE: **Long-term miglustat therapy in children with Niemann-Pick disease type C.** *J Child Neurol* 2010, **25**:300-305.
116. Wraith JE, Vecchio D, Jacklin E, Abel L, Chadha-Boreham H, Luzy C, Giorgino R, Patterson MC: **Miglustat in adult and juvenile patients with Niemann-Pick disease type C: long-term data from a clinical trial.** *Mol Genet Metab* 2010, **99**:351-357.
117. Chien YH, Lee NC, Tsai LK, Huang AC, Peng SF, Chen SJ, Hwu WL: **Treatment of Niemann-Pick disease type C in two children with miglustat: initial responses and maintenance of effects over 1 year.** *J Inherit Metab Dis* 2007, **30**:826.
118. Santos ML, Raskin S, Telles DS, Lohr JA, Liberalesso PB, Vieira SC, Cordeiro ML: **Treatment of a child diagnosed with Niemann-Pick disease type C with miglustat: A case report in Brazil.** *J Inherit Metab Dis* 2008, short report online #123.
119. Pineda M, Wraith JE, Mengel E, Sedel F, Hwu WL, Rohrbach M, Bembi B, Walterfang M, Korenke GC, Marquardt T, Luzy C, Giorgino R, Patterson MC: **Miglustat in patients with Niemann-Pick disease Type C (NP-C): A multicenter observational retrospective cohort study.** *Mol Genet Metab* 2009, **98**: 243-249.
120. Pineda M, Perez-Poyato MS, O'Callaghan M, Vilaseca MA, Pocovi M, Domingo R, Portal LR, Perez AV, Temudo T, Gaspar A, Penas JJ, Roldan S, Fumero LM, de la Barca OB, Silva MT, Macias-Vidal J, Coll MJ: **Clinical experience with miglustat therapy in pediatric patients with Niemann-Pick disease type C: a case series.** *Mol Genet Metab* 2010, **99**:358-366.
121. Jacklin E, Imrie J, Jones S, Wraith E. **Review of 11 patients with NPC1 treated with miglustat.** *Mol Genet Metab* 2010, **99**:S22
122. Wraith JE, Imrie J: **New therapies in the management of Niemann-Pick type C disease: clinical utility of miglustat.** *Ther Clin Risk Manag* 2009, **5**:877-887.
123. Yanjanin NM, Velez JI, Gropman A, King K, Bianconi SE, Conley SK, Brewer CC, Solomon B, Pavan WJ, Arcos-Burgos M, Patterson MC, Porter FD: **Linear clinical progression, independent of age of onset, in Niemann-Pick disease, type C.** *Am J Med Genet B Neuropsychiatr Genet* 2009, **153B**: 132-140.
124. Wraith JE, Guffon N, Rohrbach M, Hwu WL, Korenke GC, Bembi B, Luzy C, Giorgino R, Sedel F: **Natural history of Niemann-Pick disease type C in a multicentre observational retrospective cohort study.** *Mol Genet Metab* 2009, **98**: 250-254.
125. Floyd AG, Yu QP, Piboolnurak P, Wraith E, Patterson MC, Pullman SL: **Kinematic analysis of motor dysfunction in Niemann-Pick type C.** *Clin Neurophysiol* 2007, **118**:1010-1018.
126. Hsu AW, Piboolnurak PA, Floyd AG, Yu QP, Wraith JE, Patterson MC, Pullman SL: **Spiral analysis in Niemann-Pick disease type C.** *Mov Disord* 2009, **24**: 1984-1990.
127. Klarner B, Klünemann HH, Lurding R, Aslanidis C, Rupprecht R: **Neuropsychological profile of adult patients with Niemann-Pick C1 (NPC1) mutations.** *J Inherit Metab Dis* 2007, **30**: 60-67.
128. Tedeschi G, Bonavita S, Barton NW, Betolino A, Frank JA, Patronas NJ, Alger JR, Schiffmann R: **Proton magnetic resonance spectroscopic imaging in the clinical evaluation of patients with Niemann-Pick type C disease.** *J Neurol Neurosurg Psychiatry* 1998, **65**:72-79.
129. Galanaud D, Tourbah A, Lehericy S, Leveque N, Heron B, Billette d, V, Guffon N, Feillet F, Baumann N, Vanier MT, Sedel F: **24 month-treatment with miglustat of three patients with Niemann-Pick disease type C: follow up using brain spectroscopy.** *Mol Genet Metab* 2009, **96**:55-58.
130. Scheel M, Abegg M, Lanyon LJ, Mattman A, Barton JJ: **Eye movement and diffusion tensor imaging analysis of treatment effects in a Niemann-Pick Type C patient.** *Mol Genet Metab* 2010, **99**:291-295.
131. Choudhury A, Dominguez M, Puri V, Sharma DK, Narita K, Wheatley CL, Marks DL, Pagano RE: **Rab proteins mediate Golgi transport of caveola-internalized glycosphingolipids and correct lipid trafficking in Niemann-Pick C cells.** *J Clin Invest* 2002, **109**:1541-1550.
132. Walter M, Davies JP, Ioannou YA: **Telomerase immortalization upregulates Rab9 expression and restores LDL cholesterol egress from Niemann-Pick C1 late endosomes.** *J Lipid Res* 2003, **44**:243-253.
133. Kaptzan T, West SA, Holicky EL, Wheatley CL, Marks DL, Wang T, Peake KB, Vance J, Walkley SU, Pagano RE: **Development of a Rab9 transgenic mouse and its ability to increase the lifespan of a murine model of Niemann-Pick type C disease.** *Am J Pathol* 2009, **174**:14-20.
134. Zhang M, Strnatka D, Donohue C, Hallows JL, Vincent I, Erickson RP: **Astrocyte-only Npc1 reduces neuronal cholesterol and triples life span of Npc1<sup>-/-</sup> mice.** *J Neurosci Res* 2008, **86**:2848-2856.

135. Pentchev PG, Gal AE, Booth AD, Omodeo-Sale F, Fouks J, Neumeyer BA, Quirk JM, Dawson G, Brady RO: **A lysosomal storage disorder in mice characterized by a dual deficiency of sphingomyelinase and glucocerebrosidase.** *Biochim Biophys Acta* 1980, **619**:669-679.
136. Somers KL, Royals MA, Carstea ED, Rafi MA, Wenger DA, Thrall MA: **Mutation analysis of feline Niemann-Pick C1 disease.** *Mol Genet Metab* 2003, **79**:99-103.
137. Alvarez AR, Klein A, Castro J, Cancino GI, Amigo J, Mosqueira M, Vargas LM, Yevenes LF, Bronfman FC, Zanlungo S: **Imatinib therapy blocks cerebellar apoptosis and improves neurological symptoms in a mouse model of Niemann-Pick type C disease.** *FASEB J* 2008, **22**:3617-3627.
138. Smith D, Wallom KL, Williams IM, Jeyakumar M, Platt FM: **Beneficial effects of anti-inflammatory therapy in a mouse model of Niemann-Pick disease type C1.** *Neurobiol Dis* 2009, **36**: 242-251.
139. Mellon SH, Gong W, Schonemann MD: **Endogenous and synthetic neurosteroids in treatment of Niemann-Pick Type C disease.** *Brain Res Rev* 2008, **57**:410-420.
140. Liu B, Turley SD, Burns DK, Miller AM, Repa JJ, Dietschy JM: **Reversal of defective lysosomal transport in NPC disease ameliorates liver dysfunction and neurodegeneration in the npc1-/- mouse.** *Proc Natl Acad Sci U S A* 2009, **106**:2377-2382.
141. Ward S, O'donnell P, Fernandez S, Vite CH: **2-hydroxypropyl-beta-cyclodextrin raises hearing threshold in normal cats and in cats with Niemann-Pick type C disease.** *Pediatr Res* 2010, in press
142. Peake KB, Vance JE: **Defective cholesterol trafficking in Niemann-Pick C-deficient cells.** *FEBS Lett* 2010, doi:10.1016/J.febslet.2010.04.047
143. Ory D, Porter F, Scherrer D, Lanier M, Langmade S, Molugu V, Gale S, Olzeski D, Sidhu R, Wassif C, Yanjanin N, Schaffer J: **Cholesterol oxidation products are sensitive and specific blood-based biomarkers for Niemann-Pick C1 disease.** *Mol Genet Metab* 2010, **99**: S28

**Table 1 – NP-C functional disability scale** (from [32] and [27])

<b>Ambulation</b>	<b>Score</b>	<b>Language</b>	<b>Score</b>
Normal	1	Normal	1
Autonomous ataxic gait	2	Mild dysarthria <sup>d</sup>	2
Outdoor assisted ambulation	3	Severe dysarthria <sup>e</sup>	3
Indoor assisted ambulation	4	Non-verbal communication	4
Wheelchair bound	5	Absence of communication	5

<b>Manipulation</b>		<b>Swallowing</b>	
Normal	1	Normal	1
Slight dysmetria/dystonia <sup>a</sup>	2	Occasional dysphagia	2
Mild dysmetria/dystonia <sup>b</sup>	3	Daily dysphagia	3
Severe dysmetria/dystonia <sup>c</sup>	4	NG tube or gastric button feeding	4

Abbreviation: NG, nasogastric ; <sup>a</sup> autonomous manipulation ; <sup>b</sup> requires help for tasks but able to feed self ; <sup>c</sup> requires help for all activities ; <sup>d</sup> understandable ; <sup>e</sup> only comprehensible to certain family members.

## Legends to the Figures

### Fig. 1

#### Age of onset of neurological disease vs lifespan

Study on 97 cases for whom appropriate clinical information was available from the cohort of 181 patients originating from French hospitals. Each horizontal bar depicts one patient. The green color indicates the period during which the patient did not present neurological symptoms, irrespective of the presence or absence of preexisting systemic disease. Patients who died in their first days or months of life from systemic disease (n= 19) are not shown on this graph.

### Fig. 2

#### Niemann-Pick disease type C as a neurovisceral disease

Schematic representation of the main forms of the disease, with particular emphasis on type and age of onset of first neurological symptoms

### Fig. 3

#### Laboratory diagnosis algorithm

Footnote:

This algorithm is as proposed in Wraith et al. *Mol Genet Metab* 2009, **98**:152-165 [27]

\*Sphingomyelinase deficiency (including late-onset type A) may give a dubious filipin pattern, with normal kinetics of LDL-induced cholesteryl ester formation

\*\* False positive: I-cell disease (but very different clinical features)

\*\*\*Heterozygotes may show a pattern (filipin staining and kinetics of LDL-induced cholesteryl ester formation) similar to that in “variant” patients

\*\*\*\*In many countries, *NPCI* p.P1007A or different missense mutations on codon 992 constitute the most frequent “variant” mutations

Genetic studies can also be undertaken if clinical symptoms are very suggestive of a diagnosis of NP-C, even with negative results from filipin testing.

### Fig. 4

#### Age at diagnosis vs life span

Study in a cohort of 141 patients originating from French hospitals. Each horizontal bar depicts one patient. The change of color shows at which age the diagnosis was established.

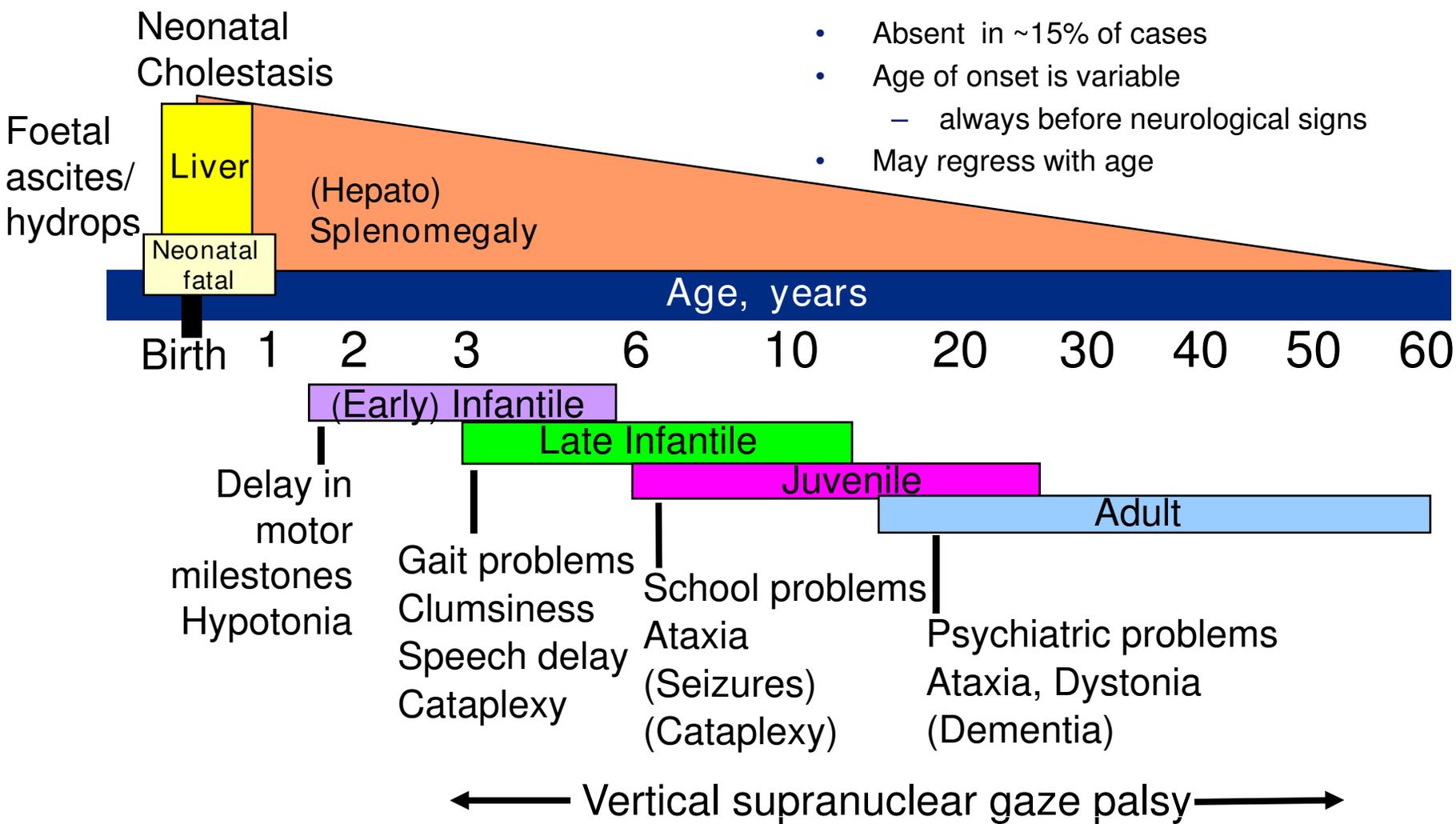
Median: 3.1 years; 0-9 months: 30%; <2 years: 36%; <5 years: 60%; > 16years: 12%



# Systemic involvement

## (hepato) Splenomegaly

- Absent in ~15% of cases
- Age of onset is variable
  - always before neurological signs
- May regress with age



# Neurological involvement

- Bone marrow (useful, not mandatory): May show foam cells (filipin + if tested for this stain)
- If a liver biopsy is performed for cholestatic liver disease, fixation for EM study is essential
- Serum chitotriosidase: useful, not mandatory; generally (not always) elevated activity
- Isolated (hepato)splenomegaly : enzymatic exclusion of Gaucher and Niemann-Pick B = prerequisite
- Provide the laboratory with sufficient clinical data (essential for correct interpretation of the results)

## SKIN BIOPSY

- If local situation permits: fixation and EM study
- Fibroblast culture (mandatory)

### FILIPIN TEST

(cell biology) (done twice)

Highly positive  
« classical »  
(85% of NP-C patients)

Moderately positive  
with pure LDL, « variant »  
(15% of NP-C patients)

Difficult Interpretation\*  
(3-5% of NP-C patients)

Clearly negative

*Nearly sure NP-C\*\**

*Probable "variant" NP-C\*\*\**

Re-assess clinical features  
*Reference Centre* Complementary  
investigations  
If likely diagnosis, gene sequencing

*a priori, not NP-C*

Kinetics of LDL-induced  
cholesteryl ester  
formation

*NPC1 Gene*  
Mutation p.P1007A  
and codon 992\*\*\*\*

### Sequencing of NPC1 and NPC2 genes

- Depending on countries, study first *NPC1* p. I1061T or other most prevalent common mutation
- Conclude quickly on *NPC2* if child < 8-10 months
- *NPC1*: numerous polymorphisms!!! – check allele segregation from parental study
- often need to study both gDNA and cDNA

Figure 4

Patients (n=141)

